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Requestor's Name: S. Devi Serial Number: 09/077.572  
Date: 24 Feb 99 Phone: 308-9347 Art Unit: 1641 (7E5)

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Please perform a search on the attached claims. Keywords and examples are provided. Please include Agricultural & Veterinary databases, US & foreign patent databases, Dissertation abstracts, Inside conferences, Medline, Biosis, EMBASE, PASCAL, JCDSTE PLUS, Derwent, FEDRIP, CRIS/USDA, TOXLINE & File 53.

Inventors: MICHAEL A. APICELLA  
MELVIN G. SUNSHINE  
NA-GYONG LEE  
RASAPPA ARUMUGHAM  
BRADFORD W. GIBSON

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CPU time: _____	Type of Search	<input checked="" type="checkbox"/> APS
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Number of Databases: <u>3</u>	<input type="checkbox"/> Structure	<input type="checkbox"/> DARC/Questel
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Devi, S.  
09/077572

09/077572

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File 440:Current Contents Search(R) 1990-1999/Mar W1

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\*File 440: Records starting 1997 to 1998W3 were reloaded, please note the changed in accession numbers.

File 71:ELSEVIER BIOBASE 1994-1999/Jan W4

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Set	Items	Description
S1	74	(HTRB OR HTR(W)B) AND (GRAM(W) (NEGATIVE OR NEG) OR SALMONE-LL? OR COLI OR HAEMOPHIL? OR HEMOPHIL?)
S2	28	RD (unique items)
S3	26	S2 AND (MUTANT? ? OR MUTAGEN? OR MUTAT? OR POLYMORPH? OR P-OLY(W)MORPHI???)

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3/3,AB/1 (Item 1 from file: 440)

DIALOG(R)File 440:Current Contents Search(R)

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09785010 GENUINE ARTICLE#: 113PM NUMBER OF REFERENCES: 49

TITLE: A lethal role for lipid A in *Salmonella* infections

AUTHOR(S): Khan SA; Everest P; Servos S; Foxwell N; Zahringer U; Brade H; Rietschel ET; Dougan G; Charles IG; Maskell DJ (REPRINT)

CORPORATE SOURCE: UNIV CAMBRIDGE,DEPT CLIN VET MED, CTR VET SCI, MADINGLEY RD/CAMBRIDGE CB3 0ES//ENGLAND/ (REPRINT); UNIV CAMBRIDGE,DEPT CLIN VET MED, CTR VET SCI/CAMBRIDGE CB3 0ES//ENGLAND/; UNIV LONDON IMPERIAL COLL

Searcher : Shears 308-4994

-key terms

09/077572

SCI TECHNOL & MED, DEPT BIOCHEM/LONDON SW7 2AY//ENGLAND/; UNIV LONDON  
UNIV COLL, RAYNE INST, WOLFSON INST BIOMED RES/LONDON WC1E 6JJ//ENGLAND/  
; BORSTEL RES CTR, CTR MED & BIOSCI/D-23845 BORSTEL//GERMANY/

PUBLICATION TYPE: JOURNAL

PUBLICATION: MOLECULAR MICROBIOLOGY, 1998, V29, N2 (JUL), P571-579

PUBLISHER: BLACKWELL SCIENCE LTD, P O BOX 88, OSNEY MEAD, OXFORD OX2 ONE,  
OXON, ENGLAND

ISSN: 0950-382X

LANGUAGE: English DOCUMENT TYPE: ARTICLE

ABSTRACT: *Salmonella* infections in naturally susceptible mice grow rapidly, with death occurring only after bacterial numbers in vivo have reached a high threshold level, commonly called the lethal load. Despite much speculation, no direct evidence has been available to substantiate a role for any candidate bacterial components in causing death. One of the most likely candidates for the lethal toxin in salmonellosis is endotoxin, specifically the lipid A domain of the lipopolysaccharide (LPS) molecule. Consequently, we have constructed a *Salmonella* mutant with a deletion-insertion in its waaN gene, which encodes the enzyme that catalyses one of the two secondary acylation reactions that complete lipid A biosynthesis. The mutant biosynthesizes a lipid A molecule lacking a single fatty acyl chain and is consequently less able to induce cytokine and inducible nitric oxide synthase (iNOS) responses both in vivo and in vitro. The mutant bacteria appear healthy, are not sensitive to increased growth temperature and synthesize a full-length O-antigen-containing LPS molecule lacking only the expected secondary acyl chain. On intravenous inoculation into susceptible BALB/c mice, wild-type *salmonellae* grew at the expected rate of approximately 10-fold per day in livers and spleens and caused the death of the infected mice when lethal loads of approximately 10<sup>8</sup> were attained in these organs. Somewhat unexpectedly, waaN mutant bacteria grew at exactly the same rate as wild-type bacteria in BALB/c mice but, when counts reached 10<sup>8</sup> per organ, mice infected with mutant bacteria survived. Bacterial growth continued until unprecedentedly high counts of 10<sup>9</sup> per organ were attained, when approximately 10% of the mice died. Most of the animals carrying these high bacterial loads survived, and the bacteria were slowly cleared from the organs. These experiments provide the first direct evidence that death in a mouse typhoid infection is directly dependent on the toxicity of lipid A and suggest that this may be mediated via proinflammatory cytokine and/or iNOS responses.

ISSN: 0950-382X

3/3, AB/2 (Item 2 from file: 440)  
DIALOG(R) File 440: Current Contents Search(R)  
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09654995 GENUINE ARTICLE#: ZZ982 NUMBER OF REFERENCES: 56  
Searcher : Shears 308-4994

TITLE: Role of the O-antigen of lipopolysaccharide, and possible roles of growth rate and of NADH:Ubiquinone oxidoreductase (nuo) in competitive tomato root-tip colonization by *Pseudomonas fluorescens* WCS365

AUTHOR(S): Dekkers LC (REPRINT); vanderBij AJ; Mulders IHM; Phoelich CC; Wentwoord RAR; Glandorf DCM; Wijffelman CA; Lugtenberg BJJ

CORPORATE SOURCE: LEIDEN UNIV, INST MOL PLANT SCI, CLUSIUS LAB, WASSENAARSEWEG 64/NL-2333 AL LEIDEN//NETHERLANDS/ (REPRINT); UNIV UTRECHT, DEPT PLANT ECOL & EVOLUTIONARY BIOL/NL-3508 TB UTRECHT//NETHERLANDS/

PUBLICATION TYPE: JOURNAL

PUBLICATION: MOLECULAR PLANT-MICROBE INTERACTIONS, 1998, V11, N8 (AUG), P 763-771

PUBLISHER: AMER PHYTOPATHOLOGICAL SOC, 3340 PILOT KNOB ROAD, ST PAUL, MN 55121

ISSN: 0894-0282

LANGUAGE: English DOCUMENT TYPE: ARTICLE

ABSTRACT: Colonization-defective, transposon-induced mutants of the efficient root colonizer *Pseudomonas fluorescens* WCS365 were identified with a gnotobiotic system. Most mutants were impaired in known colonization traits, i.e., prototrophy for amino acids, motility, and synthesis of the O-antigen of LPS (lipopolysaccharide). Mutants lacking the O-antigen of LPS were impaired in both colonization and competitive growth whereas one mutant (PCL1205) with a shorter O-antigen chain was defective only in colonization ability, suggesting a role for the intact O-antigen of LPS in colonization. Eight competitive colonization mutants that were not defective in the above-mentioned traits colonized the tomato root tip well when inoculated alone, but were defective in competitive root colonization of tomato, radish, and wheat, indicating they contained mutations affecting host range. One of these eight mutants (PCL1201) was further characterized and contains a mutation in a gene that shows homology to the *Escherichia coli* nuo4 gene, which encodes a subunit of one of two known NADH:ubiquinone oxidoreductases. Competition experiments in an oxygen-poor medium between mutant PCL1201 and its parental strain showed a decreased growth rate of mutant PCL1201. The requirement of the nuo4 gene homolog for optimal growth under conditions of oxygen limitation suggests that the root-tip environment is micro-aerobic. A mutant characterized by a slow growth rate (PCL1216) was analyzed further and contained a mutation in a gene with similarity to the *E. coli* HtrB protein, a lauroyl transferase that functions in lipid A biosynthesis.

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3/3,AB/3 (Item 3 from file: 440)  
DIALOG(R)File 440:Current Contents Search(R)  
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09/077572

09475100 GENUINE ARTICLE#: ZN277 NUMBER OF REFERENCES: 53

TITLE: Function of *Escherichia coli* MsbA, an essential ABC family transporter, in lipid A and phospholipid biosynthesis

AUTHOR(S): Zhou ZM; White KA; Polissi A; Georgopoulos C; Raetz CRH (REPRINT)

CORPORATE SOURCE: DUKE UNIV, MED CTR, DEPT BIOCHEM, POB 3711/DURHAM/NC/27710 (REPRINT); DUKE UNIV, MED CTR, DEPT BIOCHEM/DURHAM/NC/27710; CTR MED UNIV GENEVA, DEPT BIOCHIM MED/CH-1211 GENEVA 4//SWITZERLAND/

PUBLICATION TYPE: JOURNAL

PUBLICATION: JOURNAL OF BIOLOGICAL CHEMISTRY, 1998, V273, N20 (MAY 15), P 12466-12475

PUBLISHER: AMER SOC BIOCHEMISTRY MOLECULAR BIOLOGY INC, 9650 ROCKVILLE PIKE, BETHESDA, MD 20814

ISSN: 0021-9258

LANGUAGE: English DOCUMENT TYPE: ARTICLE

ABSTRACT: The *Escherichia coli* msbA gene, first identified as a multicopy suppressor of *htrB* mutations, has been proposed to transport nascent core-lipid A molecules across the inner membrane (Polissi, K., and Georgopoulos, C. (1996) Mol. Microbiol. 20, 1221-1233), msbA is an essential *E. coli* gene with high sequence similarity to mammalian Mdr proteins and certain types of bacterial ABC transporters, *htrB* is required for growth above 32 degrees C and encodes the lauroyltransferase that acts after Kdo addition during lipid A biosynthesis (Clementz, T., Bednarski, J., and Raetz, C. R. H. (1996) J. Biol. Chem. 271, 12095-12102). By using a quantitative new P-32(i) labeling technique, we demonstrate that hexa-acylated species of lipid A predominate in the outer membranes of wild type *E. coli* labeled for several generations at 42 degrees C. In contrast, in *htrB* mutants shifted to 42 degrees C for 3 h, tetraacylated lipid A species and glycerophospholipids accumulate in the inner membrane. Extra copies of the cloned msbA gene restore the ability of *htrB* mutants to grow at 42 degrees C, but they do not increase the extent of lipid A acylation. However, a significant fraction of the tetraacylated lipid A species that accumulate in *htrB* mutants are transported to the outer membrane in the presence of extra copies of msbA. *E. coli* strains in which msbA synthesis is selectively shut off at 42 degrees C accumulate hexa-acylated lipid A and glycerophospholipids in their inner membranes. Our results support the view that MsbA plays a role in lipid A and possibly glycerophospholipid transport. The tetra-acylated lipid A precursors that accumulate in *htrB* mutants may not be transported as efficiently by MsbA as are penta- or hexaacylated lipid A species.

ISSN: 0021-9258

3/3, AB/4 (Item 4 from file: 440)

DIALOG(R) File 440:Current Contents Search(R)

Searcher : Shears 308-4994

09/077572

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09035667 GENUINE ARTICLE#: YL266 NUMBER OF REFERENCES: 23

TITLE: Temperature-sensitive lesions in the *Francisella novicida* valA gene cloned into an *Escherichia coli* msbA lpxK mutant affecting deoxycholate resistance and lipopolysaccharide assembly at the restrictive temperature

AUTHOR(S): McDonald MK; Cowley SC (REPRINT); Nano FE

CORPORATE SOURCE: UNIV VICTORIA, DEPT BIOCHEM & MICROBIOL, PETCH BLDG/VICTORIA/BC V8W 3P6/CANADA/ (REPRINT); UNIV VICTORIA, DEPT BIOCHEM & MICROBIOL/VICTORIA/BC V8W 3P6/CANADA/

PUBLICATION TYPE: JOURNAL

PUBLICATION: JOURNAL OF BACTERIOLOGY, 1997, V179, N24 (DEC), P7638-7643

PUBLISHER: AMER SOC MICROBIOLOGY, 1325 MASSACHUSETTS AVENUE, NW, WASHINGTON, DC 20005-4171

ISSN: 0021-9193

LANGUAGE: English DOCUMENT TYPE: ARTICLE

ABSTRACT: The valAB locus of *Francisella novicida* has previously been found to be highly similar at the deduced amino acid level to msbA lpxK of *Escherichia coli*. Both ValA and MsbA are members of the superfamily of ABC transporters, and they appear to have similar functions. In this study we describe the isolation of a temperature-sensitive valAB locus. DNA sequence analysis indicates that the only changes to the ValAB deduced amino acid sequence are changes of S453 to an F and T458 to an I in ValA. *E. coli* strains defective in msbA and expressing temperature-sensitive ValA rapidly ceased growth when shifted from a permissive temperature to a restrictive temperature. After 1 h at the restrictive temperature, cells were much more sensitive to deoxycholate treatment. To test the hypothesis that ValA is responsible for the transport or assembly of lipopolysaccharide, we introduced gseA, a Kdo (3-deoxy-D-manno-octulosonic acid) transferase from *Chlamydia trachomatis*, into a strain with a temperature-sensitive valA allele and a nonfunctional msbA locus. These recombinants were defective in cell surface expression of the chlamydial genus-specific epitope within 15 min of a shift to the nonpermissive temperature. Also, there was enhanced association of the epitope with the inner membrane after a shift to the nonpermissive temperature. Thus, we propose that ValA is involved in the transport of lipopolysaccharide to the outer membrane.

ISSN: 0021-9193

3/3,AB/5 (Item 5 from file: 440)

DIALOG(R) File 440:Current Contents Search(R)

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08936093 GENUINE ARTICLE#: YD176 NUMBER OF REFERENCES: 70

TITLE: Study of the role of the htrB gene in *Salmonella typhimurium* virulence

Searcher : Shears 308-4994

09/077572

AUTHOR(S): Jones BD (REPRINT); Nichols WA; Gibson BW; Sunshine MG; Apicella MA

CORPORATE SOURCE: UNIV IOWA, COLL MED, DEPT MICROBIOL/IOWA CITY//IA/52242 (REPRINT); UNIV CALIF SAN FRANCISCO, SCH PHARM, DEPT PHARMACEUT CHEM/SAN FRANCISCO//CA/94143

PUBLICATION TYPE: JOURNAL

PUBLICATION: INFECTION AND IMMUNITY, 1997, V65, N11 (NOV), P4778-4783

PUBLISHER: AMER SOC MICROBIOLOGY, 1325 MASSACHUSETTS AVENUE, NW, WASHINGTON, DC 20005-4171

ISSN: 0019-9567

LANGUAGE: English DOCUMENT TYPE: ARTICLE

ABSTRACT: We have undertaken a study to investigate the contribution of the **htrB** gene to the virulence of pathogenic **Salmonella typhimurium**. An **htrB::mini-Tn10** mutation from *Escherichia coli* was transferred by transduction to the mouse-virulent strain *S. typhimurium* SL1344 to create an **htrB** mutant. The *S. typhimurium* **htrB** mutant was inoculated into mice and found to be severely limited in its ability to colonize organs of the lymphatic system and to cause systemic disease in mice. A variety of experiments were performed to determine the possible reasons for this loss of virulence. Serum killing assays revealed that the *S. typhimurium* **htrB** mutant was as resistant to killing by complement as the wild-type strain. However, macrophage survival assays revealed that the *S. typhimurium* **htrB** mutant was more sensitive to the intracellular environment of murine macrophages than the wild-type strain. In addition, the bioactivity of the lipopolysaccharide (LPS) of the **htrB** mutant was reduced compared to that of the LPS from the parent strain as measured by both a *Limulus* amoebocyte lysate endotoxin quantitation assay and a tumor necrosis factor alpha bioassay. These results indicate that the **htrB** gene plays a role in the virulence of *S. typhimurium*.

ISSN: 0019-9567

3/3,AB/6 (Item 6 from file: 440)

DIALOG(R)File 440:Current Contents Search(R)

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08936045 GENUINE ARTICLE#: YD176 NUMBER OF REFERENCES: 30

TITLE: Evaluation of the virulence of nontypeable **Haemophilus influenzae** lipooligosaccharide **htrB** and **rfaD** mutants in the chinchilla model of otitis media

AUTHOR(S): DeMaria TF (REPRINT); Apicella MA; Nichols WA; Leake ER

CORPORATE SOURCE: OHIO STATE UNIV, COLL MED, DIV OTOL RES, ROOM 4331 UHC, 456 W 10TH AVE/COLUMBUS//OH/43210 (REPRINT); UNIV IOWA, /IOWA CITY//IA/

PUBLICATION TYPE: JOURNAL

PUBLICATION: INFECTION AND IMMUNITY, 1997, V65, N11 (NOV), P4431-4435

PUBLISHER: AMER SOC MICROBIOLOGY, 1325 MASSACHUSETTS AVENUE, NW, WASHINGTON, DC 20005-4171

Searcher : Shears 308-4994

ISSN: 0019-9567

LANGUAGE: English DOCUMENT TYPE: ARTICLE

ABSTRACT: Considerable evidence has implicated nontypeable

*Haemophilus influenzae* (NTHi) lipooligosaccharide (LOS) in the pathogenesis of otitis media (OM); however, its exact role has not been conclusively established. Recently, two NTHi LOS-deficient mutants have been created and described. Strain 2019-DK1, an *rfaD* gene mutant, expresses a truncated LOS consisting of only three deoxy-D-manno-octulosonic acid residues, a single heptose, and lipid A. Strain 2019-B29, an isogenic *htrB* mutant, possesses an altered oligosaccharide core and an altered lipid A. Each strain's ability to colonize the nasopharynx and to induce OM subsequent to transbullar inoculation was evaluated in the chinchilla model. Nasopharyngeal colonization data indicate that the parent strain and both mutants are able to colonize the nasopharynx and exhibit comparable clearance kinetics. Compared with the parent and each other, however, the mutants demonstrated marked differences in virulence regarding their relative abilities to induce OM and persist in the middle ear post-transbullar inoculation. Strain B29 required a 3-log-greater dose to induce OM than the parent strain and did not exhibit evidence of sustained multiplication but persisted for the same duration as the parent. Conversely, strain-DK1, even when inoculated at a dose 4 logs greater than the parent dose, was eliminated from the middle ear 72 h after challenge. A comparison of the relative pathogenicities of these isolates provides the opportunity to address fundamental questions regarding the contribution of LOS to pathogenesis issues at the molecular level. Specifically, the impact of these LOS gene disruptions on OM pathogenesis can be defined and may thus provide potential new targets for future protection and intervention strategies.

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3/3,AB/7 (Item 7 from file: 440)

DIALOG(R)File 440:Current Contents Search(R)

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08773455 GENUINE ARTICLE#: XT829 NUMBER OF REFERENCES: 28

TITLE: *htrB* of *Haemophilus influenzae*: determination of biochemical activity and effects on virulence and lipooligosaccharide toxicity

AUTHOR(S): Nichols WA; Raetz CRH; Clementz T; Smith AL; Hanson JA; Ketterer MR; Sunshine M; Apicella MA (REPRINT)

CORPORATE SOURCE: UNIV IOWA, COLL MED, DEPT MICROBIOL BSB 3 403, 51 NEWTON RD/IOWA CITY//IA/52242 (REPRINT); UNIV IOWA, COLL MED, DEPT MICROBIOL BSB 3 403/IOWA CITY//IA/52242; DUKE UNIV, MED CTR, DEPT BIOCHEM/DURHAM//NC/27710; UNIV MISSOURI, SCH MED, DEPT MOL MICROBIOL & IMMUNOL/COLUMBIA//MO/65212

PUBLICATION TYPE: JOURNAL

Searcher : Shears 308-4994



PUBLICATION: JOURNAL OF ENDOTOXIN RESEARCH, 1997, V4, N3 (JUN), P163-172  
 PUBLISHER: CHURCHILL LIVINGSTONE, JOURNAL PRODUCTION DEPT, ROBERT STEVENSON  
 HOUSE, 1-3 BAXTERS PLACE, LEITH WALK, EDINBURGH EH1 3AF, MIDLOTHIAN,  
 SCOTLAND

ISSN: 0968-0519

LANGUAGE: English DOCUMENT TYPE: ARTICLE

ABSTRACT: The *htrB* mutant of *Haemophilus influenzae*

(strain B29) has been shown to lack secondary (nonhydroxylated) acyl groups in its lipid A. We have determined through in vitro biochemical assays that the *HtrB* protein acts as a specific acyltransferase in the late stages of lipid A biosynthesis and that the preferred acyl group donor is myristoyl-acyl carrier protein. Under the conditions employed, the *Escherichia coli* precursor, Kdo(2)-lipid IVA, functions as a myristate acceptor. Introduction of the *Haemophilus htrB* gene into an *E. coli* mutant lacking *htrB* complements the biochemical and physiological defects associated with the *E. coli htrB* mutation.

Tumor necrosis factor alpha (TNF alpha) assays using murine and human macrophage cells indicated that nontypeable *H. influenzae* (NtHi) strain 2019 and *H. influenzae* type b strain A2 elicit levels of expression of TNF alpha that are 30-40 times greater than levels induced by the isogenic *htrB* mutants (B29 and A2B29). Studies using cell-free LOS indicated that the LOS from wild type strain 2019 elicits levels of TNF alpha expression that are 6-8-fold higher than those of B29. In situ hybridization studies of a primary human bronchial epithelial cell line demonstrated a greater increase of TNF alpha message produced in the presence of 2019 LOS than in the presence of B29 LOS. TNF alpha levels of the cell supernatant of cells stimulated with 2019 LOS were found to be 7-8-fold higher than levels in B29 stimulated supernatants. Using the *Limulus* amoebocyte lysate for assessment of endotoxic activity, we found that wild type LOS was 8-fold higher in endotoxic activity compared with the mutant LOS. In virulence assays using intraperitoneal inoculation of infant rats, the *htrB* isogenic strain caused bacteremia at 50% the frequency of the wild type strain. In intranasal inoculation studies, the *htrB* mutant strain was unable to cause bacteremia whereas the wild type b parent produced bacteremia in 40-60% of the animals. These findings suggest that the *htrB* gene of *H. influenzae* is important for virulence and that host TNF alpha expression is attenuated in response to *htrB* mutant strains.

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3/3,AB/8 (Item 8 from file: 440)  
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08773215 GENUINE ARTICLE#: XT850 NUMBER OF REFERENCES: 69  
 Searcher : Shears 308-4994

TITLE: Identification of the gene encoding the *Escherichia coli* lipid  
a 4'-kinase - Facile phosphorylation of endotoxin analogs with  
recombinant LpxK

AUTHOR(S): Garrett TA; Kadrmaz JL; Raetz CRH (REPRINT)

CORPORATE SOURCE: DUKE UNIV, MED CTR, DEPT BIOCHEM/DURHAM//NC/27710  
(REPRINT); DUKE UNIV, MED CTR, DEPT BIOCHEM/DURHAM//NC/27710

PUBLICATION TYPE: JOURNAL

PUBLICATION: JOURNAL OF BIOLOGICAL CHEMISTRY, 1997, V272, N35 (AUG 29), P  
21855-21864

PUBLISHER: AMER SOC BIOCHEMISTRY MOLECULAR BIOLOGY INC, 9650 ROCKVILLE  
PIKE, BETHESDA, MD 20814

ISSN: 0021-9258

LANGUAGE: English DOCUMENT TYPE: ARTICLE

ABSTRACT: The genes for seven of nine enzymes needed for the biosynthesis  
of Kdo(2)-lipid A (Re endotoxin) in *Escherichia coli* have been  
reported. We have now identified a novel gene encoding the lipid A  
4'-kinase (the sixth step of the pathway). The 4'-kinase transfers the  
(gamma) over dot-phosphate of ATP to the 4'-position of a  
tetraacyldisaccharide 1-phosphate intermediate (termed DS-1-P) to form  
tetraacyldisaccharide 1,4'-bis-phosphate (lipid IVA). The 4'-phosphate  
is required for the action of distal enzymes, such as Kdo transferase  
and also renders lipid A substructures active as endotoxin antagonists  
or mimetics. Lysates of *E. coli* generated using individual A  
clones from the ordered Kohara library were assayed for over-production  
of 4'-kinase. Only one clone, [218]E1D1, which directed 2-2.5-fold  
overproduction, was identified. This construct contains 20 kilobase  
pairs of *E. coli* DNA from the vicinity of minute 21. Two genes  
related to the lipid A system map in this region: msbA, encoding a  
putative translocator, and kdsB, the structural gene for CMP-Kdo  
synthase, msbA forms an operon with a downstream, essential open  
reading frame of unknown function, designated orfE. orfE was cloned  
into a T7 expression system. Washed membranes from cells overexpressing  
orfE display similar to 2000-fold higher specific activity of 4'-kinase  
than membranes from cells with vector alone. Membranes containing  
recombinant, overexpressed 4'-kinase (but not membranes with wild-type  
kinase levels) efficiently phosphorylate three DS-1-P analogs:  
3-aza-DS-1-P, base-treated DS-1-P, and base-treated 3-aza-DS-1-P. A  
synthetic hexaacylated DS-1-P analog, compound 505, can also be  
phosphorylated by membranes from the overproducer, yielding [4'-P-32]  
lipid A (endotoxin). The overexpressed lipid A 4'-kinase is very useful  
for making new 4'-phosphorylated lipid A analogs with potential utility  
as endotoxin mimetics or antagonists. We suggest that orfE is the  
structural gene for the 4'-kinase and that it be redesignated IpxK.

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3/3,AB/9 (Item 9 from file: 440)

DIALOG(R)File 440:Current Contents Search(R)

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Searcher : Shears 308-4994

08764778 GENUINE ARTICLE#: XT772 NUMBER OF REFERENCES: 36

TITLE: Mutation of the *htrB* gene in a virulent *Salmonella* typhimurium strain by intergeneric transduction: Strain construction and phenotypic characterization

AUTHOR(S): Sunshine MG; Gibson BW; Engstrom JJ; Nichols WA; Jones BD; Apicella MA (REPRINT)

CORPORATE SOURCE: UNIV IOWA, COLL MED, DEPT MICROBIOL, BSB 3-403, 51 NEWTON RD/IOWA CITY//IA/52242 (REPRINT); UNIV IOWA, COLL MED, DEPT MICROBIOL/IOWA CITY//IA/52242; UNIV CALIF SAN FRANCISCO, SCH PHARM, DEPT PHARMACEUT CHEM/SAN FRANCISCO//CA/94143

PUBLICATION TYPE: JOURNAL

PUBLICATION: JOURNAL OF BACTERIOLOGY, 1997, V179, N17 (SEP), P5521-5533

PUBLISHER: AMER SOC MICROBIOLOGY, 1325 MASSACHUSETTS AVENUE, NW, WASHINGTON, DC 20005-4171

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LANGUAGE: English DOCUMENT TYPE: ARTICLE

ABSTRACT: The *htrB* gene product of *Haemophilus influenzae* contributes to the toxicity of the lipooligosaccharide. The *htrB* gene encodes a 2-keto-3-deoxyoctulosonic acid-dependent acyltransferase which is responsible for myristic acid substitutions at the hydroxy moiety of lipid A beta-hydroxymyristic acid. Mass spectroscopic analysis has demonstrated that lipid A from an *H. influenzae htrB* mutant is predominantly tetraacyl and similar in structure to lipid TV,, which has been shown to be nontoxic in animal models. We sought to construct a *Salmonella typhimurium htrB* mutant in order to investigate the contribution of *htrB* to virulence in a well-defined murine typhoid model of animal pathogenesis. To this end, all *r*(-) *m*(+) *galE* *mutS* *recD* strain of *S. typhimurium* was constructed (MGS-7) and used in inter- and intrastain transduction experiments with both coliphage P1 and *Salmonella* phage P22. The *Escherichia coli htrB* gene containing a mini-Tn10 insertion was transduced from *E. coli* MLK217 into *S. typhimurium* MGS-7 via phage P1 and subsequently via phage P22 into the virulent-*Salmonella* strain SL1344. All *S. typhimurium* transductants showed phenotypes similar to those described for the *E. coli htrB* mutant. Mass spectrometric analysis of the crude lipid A fraction from the lipopolysaccharide of the *S. typhimurium htrB* mutant strain showed that for the dominant hexaacyl form, a lauric acid moiety was lost at one position on the lipid A and a palmitic acid moiety was added at another position; for the less abundant heptaacyl species, the lauric acid was replaced with palmitoleic acid.

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3/3,AB/10 (Item 10 from file: 440)

DIALOG(R) File 440:Current Contents Search(R)

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Searcher : Shears 308-4994

08467502 GENUINE ARTICLE#: WZ944 NUMBER OF REFERENCES: 36

TITLE: The outer membrane of lipid A-deficient *Escherichia coli* mutant LH530 has reduced levels of OmpF and leaks periplasmic enzymes

AUTHOR(S): Nurminen M (REPRINT); Hirvas L; Vaara M

CORPORATE SOURCE: UNIV HELSINKI, DEPT BACTERIOL & IMMUNOL, HAARTMAN INST, HAARTMANINKATU 3, POB 21/SF-00014 HELSINKI//FINLAND/ (REPRINT)

PUBLICATION TYPE: JOURNAL

PUBLICATION: MICROBIOLOGY-UK, 1997, V143, ,5 (MAY), P1533-1537

PUBLISHER: SOC GENERAL MICROBIOLOGY, MARLBOROUGH HOUSE, BASINGSTOKE RD, SPENCERS WOODS, READING, BERKS, ENGLAND RG7 1AE

ISSN: 1350-0872

LANGUAGE: English DOCUMENT TYPE: ARTICLE

ABSTRACT: We have previously described a new *Escherichia coli* K-12 mutant, LH530, which has a defective outer membrane. LH530 is very sensitive to hydrophobic antibiotics, does not grow at 42 degrees C and synthesizes reduced amounts of lipid A. Phenotypically LH530 is very similar to the known lipid A biosynthesis mutants of *E. coli* and *Salmonella typhimurium*. Its genetic defect is not known, but the defect is suppressed by multiple copies of ORF195. Here we show that at 37 degrees C LH530 contains a reduced amount of the OmpF porin and that it leaks periplasmic beta-lactamase at 37 degrees C and 42 degrees C. We further show that ORF195, when present at low copy number, restores the antibiotic resistance and lipid A biosynthesis of LH530 at 28 degrees C. but not at higher temperatures. In contrast, OmpF expression is restored at 37 degrees C.

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3/3,AB/11 (Item 11 from file: 440)

DIALOG(R)File 440:Current Contents Search(R)

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08382629 GENUINE ARTICLE#: WV262 NUMBER OF REFERENCES: 34

TITLE: Function of the *Escherichia coli* msbB gene, a multicopy suppressor of *htrB* knockouts, in the acylation of lipid A - Acylation by MsbB follows laurate incorporation by *HtrB*

AUTHOR(S): Clementz T; Zhou ZM; Raetz CRH (REPRINT)

CORPORATE SOURCE: DUKE UNIV, MED CTR, DEPT BIOCHEM/DURHAM//NC/27710 (REPRINT); DUKE UNIV, MED CTR, DEPT BIOCHEM/DURHAM//NC/27710

PUBLICATION TYPE: JOURNAL

PUBLICATION: JOURNAL OF BIOLOGICAL CHEMISTRY, 1997, V272, N16 (APR 18), P 10353-10360

PUBLISHER: AMER SOC BIOCHEMISTRY MOLECULAR BIOLOGY INC, 9650 ROCKVILLE PIKE, BETHESDA, MD 20814

ISSN: 0021-9258

LANGUAGE: English DOCUMENT TYPE: ARTICLE

ABSTRACT: Overexpression of the *Escherichia coli* msbB gene on high Searcher : Shears 308-4994

copy plasmids suppresses the temperature-sensitive growth associated with mutations in the *htrB* gene, *htrB* encodes the lauroyl transferase of lipid A biosynthesis that acylates the intermediate (Kdo)(2)-lipid IVA (Brozek, It. A. and Raetz, C. R. H. (1990) J. Biol. Chem, 265, 15410-15417). Since *msbB* displays 27.5% identity and 42.2% similarity to *htrB*, we explored the possibility that *msbB* encodes a related acyltransferase, In contrast to *htrB*, extracts of strains with insertion mutations in *msbB* are not defective in transferring laurate from lauroyl acyl carrier protein to (Kdo)(2)-lipid IVA. However, extracts of *msbB* mutants do not efficiently acylate the product formed by *HtrB*, designated (IZdo)(2)-(lauroyl)-lipid IVA. Extracts of strains harboring *msbB*(+) bearing plasmids acylate (Kdo)(2)-(lauroyl)-lipid IVA very rapidly compared with wild type, We solubilized and partially purified *MsbB* from an overproducing strain, lacking *HtrB*, *MsbB* transfers myristate or laurate, activated on ACP, to (Kdo)(2)-(lauroyl)-lipid IVA. Decanoyl, palmitoyl, palmitoleoyl, and (R)-3-hydroxymyristoyl-ACP are poor acyl donors, *MsbB* acylates (Kdo)(2)-(lauroyl)-lipid IVA about 100 times faster than (Kdo)(2)-lipid IVA. The slow, but measurable, rate whereby *MsbB* acts on (Kdo)(2)-lipid IVA may explain why overexpression of *MsbB* suppresses the temperature-sensitive phenotype of *htrB* mutations, Presumably, the acyloxyacyl group generated by excess *MsbB* substitutes for the one normally formed by *HtrB*.

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3/3,AB/12 (Item 12 from file: 440)  
 DIALOG(R)File 440:Current Contents Search(R)  
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08131494 GENUINE ARTICLE#: WE411 NUMBER OF REFERENCES: 52

TITLE: The lipid A biosynthesis deficiency of the *Escherichia coli* antibiotic-supersensitive mutant LH530 is suppressed by a novel locus, ORF195

AUTHOR(S): Hirvas L (REPRINT); Nurminen M; Helander IM; Vuorio R; Vaara M  
 CORPORATE SOURCE: UNIV HELSINKI, HAARTMAN INST, DEPT BACTERIOL & IMMUNOL,  
 POB 21, HAARTMANINKATU 3/SF-00014 HELSINKI//FINLAND/ (REPRINT); NATL  
 PUBL HLTH INST, DEPT BACTERIAL VACCINE RES & MOL BIOL/FIN-00300  
 HELSINKI//FINLAND/

PUBLICATION TYPE: JOURNAL

PUBLICATION: MICROBIOLOGY-UK, 1997, V143, ,1 (JAN), P73-81

PUBLISHER: SOC GENERAL MICROBIOLOGY, MARLBOROUGH HOUSE, BASINGSTOKE RD,  
 SPENCERS WOODS, READING, BERKS, ENGLAND RG7 1AE

ISSN: 1350-0872

LANGUAGE: English DOCUMENT TYPE: ARTICLE

ABSTRACT: A new mutant of *Escherichia coli* K-12 supersensitive to both hydrophobic and large hydrophilic antibiotics was isolated and characterized. The mutant grew well at 28 degrees C, poorly at 37

Searcher : Shears 308-4994

degrees C. and did not grow at 42 degrees C. The rate of its lipid A biosynthesis was reduced as compared to that of the parent strain. This deficiency was rescued by a novel locus, ORF195, the function of which has not been elucidated. ORF195 is located in the 76 min region in the *E. coli* chromosome and encodes a hypothetical 21 <bulet>8 kDa protein with no signal sequence. ORF195 isolated from the mutant strain had an identical sequence to the wild-type allele, indicating a suppressor function of the gene product.

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3/3,AB/13 (Item 13 from file: 440)  
 DIALOG(R)File 440:Current Contents Search(R)  
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07517367 GENUINE ARTICLE#: UU880 NUMBER OF REFERENCES: 43  
 TITLE: **MUTATIONAL ANALYSIS AND PROPERTIES OF THE MSBA GENE OF**  
**ESCHERICHIA COLI, CODING FOR AN ESSENTIAL ABC FAMILY TRANSPORTER**  
 AUTHOR(S): POLISSI A; GEORGOPOULOS C  
 CORPORATE SOURCE: GLAXO RIC,DIV MICROBIOL,VIA FLEMING 2/I-37100  
 VERONA//ITALY/ (Reprint); CTR MED UNIV GENEVA,DEPT BIOCHIM MED/CH-1211  
 GENEVA 4//SWITZERLAND/

PUBLICATION: MOLECULAR MICROBIOLOGY, 1996, V20, N6 (JUN), P1221-1233

ISSN: 0950-382X

LANGUAGE: ENGLISH DOCUMENT TYPE: ARTICLE

ABSTRACT: The *htrB* gene was discovered because its insertional inactivation interfered with *Escherichia coli* growth and viability at temperatures above 32.5 degrees C, as a result of accumulation of phospholipids. The *msbA* gene was originally discovered because when cloned on a low-copy-number plasmid vector it was able to suppress the temperature-sensitive growth phenotype of an *htrB* null mutant as well as the accumulation of phospholipids. The *msbA* gene product belongs to the superfamily of ABC transporters, a universally conserved family of proteins characterized by a highly conserved ATP-binding domain. The *msbA* gene is essential for bacterial viability at all temperatures. In order to understand the physiological role of the MsbA protein, we mutated the ATP-binding domain using random PCR mutagenesis. Six independent mutants were isolated and characterized. Four of these mutations resulted in single-amino-acid substitutions in non-conserved residues and were able to support cell growth at 30 degrees C but not at 43 degrees C. The remaining two mutations behaved as recessive lethals, and resulted in single-amino-acid substitutions in Walker motif B, one of the two highly conserved regions of the ATP-binding domain. Despite the fact that neither of these two mutant proteins can support *E. coli* growth, they both retained the ability to bind ATP in vitro. In addition, we present evidence to show that N-acetyl [H-3]-glucosamine, a precursor of lipopolysaccharides, accumulates at the non-permissive temperature in the inner membrane of either *htrB* null or *msbA*

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conditional lethal strains, Translocation of the precursor to the outer membrane is restored by transformation with a plasmid containing the wild-type *msbA* gene, A possible role for *MsbA* as a translocator of lipopolysaccharides or its precursors is discussed.

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3/3,AB/14 (Item 14 from file: 440)  
DIALOG(R)File 440:Current Contents Search(R)  
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07378476 GENUINE ARTICLE#: UL250 NUMBER OF REFERENCES: 56

TITLE: FUNCTION OF THE *htrB* HIGH TEMPERATURE REQUIREMENT GENE OF  
*ESCHERICHIA COLI* IN THE ACYLATION OF LIPID A - *htrB*  
CATALYZED INCORPORATION OF LAURATE

AUTHOR(S): CLEMENTZ T; BEDNARSKI JJ; RAETZ CRH (Reprint)

CORPORATE SOURCE: DUKE UNIV, MED CTR, DEPT BIOCHEM/DURHAM//NC/27710 (Reprint)  
; DUKE UNIV, MED CTR, DEPT BIOCHEM/DURHAM//NC/27710

PUBLICATION: JOURNAL OF BIOLOGICAL CHEMISTRY, 1996, V271, N20 (MAY 17), P  
12095-12102

ISSN: 0021-9258

LANGUAGE: ENGLISH DOCUMENT TYPE: ARTICLE

ABSTRACT: By assaying lysates of *Escherichia coli* generated with the hybrid lambda bacteriophages of an ordered library (Kohara, Y., Akiyama, K., and Isono, K. (1987) Cell 50, 495-508), we identified two clones (lambda 232 and lambda 233) capable of overexpressing the lauroyl transferase that functions after 3-deoxy-D-manno-octulosonic acid (Kdo) addition in lipid A biosynthesis (Brozek, K. A., and Raetz, C. R. H. (1990) J. Biol. Chem. 265, 15410-15417). The *E. coli* DNA inserts in lambda 232 and lambda 233 suggested that a known gene (*htrB*) required for rapid growth above 33 degrees C might encode the lauroyl transferase. Using the intermediate (Kdo)(2)-lipid IVA as the laurate acceptor, extracts of strains with transposon insertions in *htrB* were found to contain no lauroyl transferase activity. Cells harboring hybrid *htrB*(+) plasmids overproduced transferase activity 100-200-fold. The overproduced transferase was solubilized with a non-ionic detergent and purified further by DEAE-Sepharose chromatography. With lauroyl acyl carrier protein as the donor, the purified enzyme rapidly incorporated one laurate residue into (Kdo)(2)-lipid IVA. The rate of laurate incorporation was reduced by several orders of magnitude when either one or both Kdos were absent in the acceptor. With a matched set of acyl-acyl carrier proteins, the enzyme incorporated laurate 3-8 times faster than decanoate or myristate, respectively. Transfer of palmitate, palmitoleate, or R-3-hydroxymyristate was very slow. Taken together with previous studies, our findings indicate that *htrB* encodes a key, late functioning acyltransferase of lipid A biosynthesis.

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3/3,AB/15 (Item 15 from file: 440)  
 DIALOG(R)File 440:Current Contents Search(R)  
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06897494 GENUINE ARTICLE#: TE583 NUMBER OF REFERENCES: 30

TITLE: **MUTATION OF THE HTRB LOCUS OF HAEMOPHILUS**

INFLUENZAE NONTYPABLE STRAIN 2019 IS ASSOCIATED WITH MODIFICATIONS OF  
 LIPID A AND PHOSPHORYLATION OF THE LIPO-OLIGOSACCHARIDE

AUTHOR(S): LEE NG; SUNSHINE MG; ENGSTROM JJ; GIBSON BW; APICELLA  
 MA (Reprint)

CORPORATE SOURCE: UNIV IOWA,DEPT MICROBIOL,BOWEN SCI BLDG,51 NEWTON RD/IOWA  
 CITY//IA/52242 (Reprint); UNIV IOWA,DEPT MICROBIOL/IOWA CITY//IA/52242;  
 UNIV CALIF SAN FRANCISCO,SCH PHARM,DEPT PHARMACEUT CHEM/SAN  
 FRANCISCO//CA/94143

PUBLICATION: JOURNAL OF BIOLOGICAL CHEMISTRY, 1995, V270, N45 (NOV 10), P  
 27151-27159

ISSN: 0021-9258

LANGUAGE: ENGLISH DOCUMENT TYPE: ARTICLE

ABSTRACT: The **HtrB** protein was first identified in *Escherichia coli* as a protein required for cell viability at high temperature, but its expression was not regulated by temperature. We isolated an **htrB** homologue from nontypable *Haemophilus influenzae* strain (NTHi) 2019, which was able to functionally complement the *E. coli* **htrB** mutation. The promoter for the NTHi 2019 **htrB** gene overlaps the promoter for the *rfaE* gene, and the two genes are divergently transcribed. The deduced amino acid sequence of NTHi 2019 **HtrB** had 56% homology to *E. coli* **HtrB**. In vitro transcription-translation analysis confirmed production of a protein with an apparent molecular mass of 32-33 kDa. Primer extension analysis revealed that **htrB** was transcribed from a sigma(70)-dependent consensus promoter and its expression was not affected by temperature. The expression of **htrB** and *rfaE* was 2.5-4 times higher in the NTHi **htrB** mutant B29 than in the parental strain. In order to study the function of the **HtrB** protein in *Haemophilus*, we generated two isogenic **htrB** mutants by shuttle mutagenesis using a mini-Tn3. The **htrB** mutants initially showed temperature sensitivity, but they lost the sensitivity after a few passages at 30 degrees C and were able to grow at 37 degrees C. They also showed hypersensitivity to deoxycholate and kanamycin, which persisted on passage. SDS-polyacrylamide gel electrophoresis analysis revealed that the lipo-oligosaccharide (LOS) isolated from these mutants migrated faster than the wild type LOS and its color changed from black to brown as has been described for *E. coli* **htrB** mutants. Immunoblotting analysis also showed that the LOS from the **htrB** mutants lost reactivity to a monoclonal antibody, 6E4, which binds to the wild type NTHi 2019 LOS. Electrospray ionization-mass spectrometry analysis of the O-deacylated LOS oligosaccharide indicated a modification of the core structure

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characterized in part by a net loss in phosphoethanolamine. Mass spectrometric analysis of the lipid A of the **htrB** mutant indicated a loss of one or both myristic acid substitutions. These data suggest that **HtrB** is a multifunctional protein and may play a controlling role in regulating cell responses to various environmental changes.

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3/3,AB/16 (Item 16 from file: 440)  
DIALOG(R)File 440:Current Contents Search(R)  
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06192373 GENUINE ARTICLE#: QJ524 NUMBER OF REFERENCES: 36

TITLE: MOLECULAR CLONING AND CHARACTERIZATION OF THE NONTYPEABLE  
**HAEMOPHILUS INFLUENZAE** 2019 RFAE GENE REQUIRED FOR  
LIPOPOLYSACCHARIDE BIOSYNTHESIS

AUTHOR(S): LEE NG; SUNSHINE MG; APICELLA MA (Reprint)

CORPORATE SOURCE: UNIV IOWA,DEPT MICROBIOL,BOWEN SCI BLDG 3-401,51 NEWTON  
RD/IOWA CITY//IA/52242 (Reprint); UNIV IOWA,DEPT MICROBIOL/IOWA  
CITY//IA/52242

PUBLICATION: INFECTION AND IMMUNITY, 1995, V63, N3 (MAR), P818-824

ISSN: 0019-9567

LANGUAGE: ENGLISH DOCUMENT TYPE: ARTICLE

ABSTRACT: The lipooligosaccharide (LOS) of nontypeable **Haemophilus influenzae** (NTHi) is an important factor in pathogenesis and virulence. In an attempt to elucidate the genes involved in LOS biosynthesis, we have cloned the **rfaE** gene from NTHi 2019 by complementing a **Salmonella typhimurium rfaE** mutant strain with an NTHi 2019 plasmid library. The **rfaE** mutant synthesizes lipopolysaccharide (LPS) lacking heptose, and the **rfaE** gene is postulated to be involved in ADP-heptose synthesis. Retransformation with the plasmid containing 4 kb of NTHi DNA isolated from a reconstituted mutant into **rfaE** mutants gave wild-type LPS phenotypes. Sodium dodecyl sulfate-polyacrylamide gel electrophoresis analysis confirmed the conversion of the **rfaE** mutant LPS to a wild-type LPS phenotype. Sequence analysis of a 2.4-kb Bg/II fragment revealed two open reading frames. One open reading frame encodes the RfaE protein with a molecular weight of 37.6 kDa, which was confirmed by in vitro transcription and translation, and the other encodes a polypeptide highly homologous to the *Escherichia coli* **HtrB** protein. These two genes are transcribed from the same promoter region into opposite directions. Primer extension analysis of the **rfaE** gene revealed a single transcription start site at 37 bp upstream of the predicted translation start. site. The upstream promoter region contained a sequence (TA AAAT) homologous to the -10 region of the bacterial sigma(70)-dependent promoters at an appropriate distance (7 bp), but no sequence resembling the consensus sequence of the -35 region was found. These studies demonstrate the ability to use

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09/077572

complementation of defined LPS defects in members of the family  
Enterobacteriaceae to identify LOS synthesis genes in NTHi.

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3/3,AB/17 (Item 17 from file: 440)  
DIALOG(R)File 440:Current Contents Search(R)  
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06026225 GENUINE ARTICLE#: PY759 NUMBER OF REFERENCES: 44  
TITLE: SERUM-SENSITIVE **MUTATION** OF FRANCISELLA NOVICIDA - ASSOCIATION  
WITH AN ABC TRANSPORTER GENE  
AUTHOR(S): MDLULI KE; ANTHONY LSD; BARON GS; MCDONALD MK; MYLTSEVA SV; NANO  
FE  
CORPORATE SOURCE: UNIV VICTORIA,DEPT BIOCHEM & MICROBIOL/VICTORIA/BC V8W  
3P6/CANADA/ (Reprint)  
PUBLICATION: MICROBIOLOGY-UK, 1994, V140, DEC (DEC), P3309-3318  
ISSN: 1350-0872  
LANGUAGE: ENGLISH DOCUMENT TYPE: ARTICLE

ABSTRACT: Francisella novicida is a facultative intracellular pathogen that  
can survive and grow in macrophages by preventing phagolysosomal  
fusion. In this study in vitro cassette **mutagenesis** was used to  
generate a library of insertion **mutants** of F. novicida. Two  
related **mutants**, KM14 and KM14S, initially identified as  
defective for growth in macrophages, were found to be sensitive to  
serum. These **mutants** were also found to grow approximately  
1000-fold less well in the livers and spleens of infected mice. We  
cloned a genetic locus that was presumably **mutagenized** in these  
**mutants** and found that it included genes that had high similarity  
in their deduced amino acid sequence to those of msbA and orfE of  
Escherichia coli. The former is a member of the superfamily of ABC  
transporter proteins. We named the corresponding genes in F. novicida,  
valAB. Integration of a cloned valAB locus into the chromosome of KM14S  
partially restored the serum resistance phenotype found in wild-type F.  
novicida.

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3/3,AB/18 (Item 18 from file: 440)  
DIALOG(R)File 440:Current Contents Search(R)  
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04227251 GENUINE ARTICLE#: KE353 NUMBER OF REFERENCES: 40  
TITLE: THE ESSENTIAL ESCHERICHIA-COLI MSBA GENE, A MULTICOPY  
SUPPRESSOR OF NULL **MUTATIONS** IN THE HTRB GENE, IS RELATED  
TO THE UNIVERSALLY CONSERVED FAMILY OF ATP-DEPENDENT TRANSLOCATORS  
AUTHOR(S): KAROW M; GEORGOPOULOS C  
CORPORATE SOURCE: TEMPLE UNIV,HLTH SCI CTR,DEPT MICROBIOL & IMMUNOL,3400 N  
BROAD ST/PHILADELPHIA//PA/19140 (Reprint); UNIV UTAH,SCH MED,DEPT  
Searcher : Shears 308-4994

CELLULAR VIRAL & MOLEC BIOL/SALT LAKE CITY//UT/84132; CTR MED UNIV  
GENEVA/CH-1211 GENEVA 4//SWITZERLAND/

PUBLICATION: MOLECULAR MICROBIOLOGY, 1993, V7, N1 (JAN), P69-79

ISSN: 0950-382X

LANGUAGE: ENGLISH DOCUMENT TYPE: ARTICLE

ABSTRACT: We report the characterization of the *msbA* gene, isolated as a multicopy suppressor of the *HtrB* temperature-sensitive phenotype. The *msbA* gene maps to 20.5 min on the *Escherichia coli* genetic map and encodes a protein with an estimated molecular mass of 64460 Da, with the properties of an integral membrane protein. The amino acid sequence of *MsbA* is very similar to those of the family of ATP-dependent translocators, which includes the haemolysin B protein of *E. coli* and the mammalian multidrug resistance (MDR) proteins. Mutational analysis of *msbA* indicates that it may form an operon with a downstream gene, *orfE*, and that both of these genes are essential for bacterial viability under all growth conditions tested.

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3/3,AB/19 (Item 19 from file: 440)

DIALOG(R)File 440:Current Contents Search(R)

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04134812 GENUINE ARTICLE#: JY560 NUMBER OF REFERENCES: 47

TITLE: THE LETHAL PHENOTYPE CAUSED BY NULL MUTATIONS IN THE  
ESCHERICHIA-COLI-HTRB GENE IS SUPPRESSED BY MUTATIONS  
IN THE ACCBC OPERON, ENCODING 2 SUBUNITS OF ACETYL COENZYME A  
CARBOXYLASE

AUTHOR(S): KAROW M; FAYET O; GEORGOPOULOS C

CORPORATE SOURCE: TEMPLE UNIV, HLTH SCI CTR, DEPT MICROBIOL &  
IMMUNOL/PHILADELPHIA//PA/19140 (Reprint); UNIV UTAH, SCH MED, DEPT  
CELLULAR VIRAL & MOLEC BIOL/SALT LAKE CITY//UT/84132; CTR RECH BIOCHIM  
& GENET CELLULAIRES, CNRS/F-31062 TOULOUSE//FRANCE/; UNIV GENEVA, CTR  
MED/CH-1211 GENEVA 4//SWITZERLAND/

PUBLICATION: JOURNAL OF BACTERIOLOGY, 1992, V174, N22 (NOV), P7407-7418

ISSN: 0021-9193

LANGUAGE: ENGLISH DOCUMENT TYPE: ARTICLE

ABSTRACT: Insertion mutations in the *Escherichia coli*

*htrB* gene result in the unique phenotype of not affecting growth at temperatures below 32.5-degrees-C but leading to a loss of viability at temperatures above this in rich media. When *htrB* bacteria growing in rich media were shifted to the nonpermissive temperature of 42-degrees-C, they continued to grow at a rate similar to that at 30-degrees-C but they produced phospholipids at the rate required for growth at 42-degrees-C. This led to the accumulation of more than twice as much phospholipid per milligram of protein compared with that in wild-type bacteria. Consistent with *HtrB* playing a role in phospholipid biosynthesis, one complementation group of spontaneously arising mutations that suppressed *htrB*-induced lethality

Searcher : Shears 308-4994

were mapped to the accBC operon. This operon codes for the biotin carboxyl carrier protein and biotin carboxylase subunits of the acetyl coenzyme A carboxylase enzyme complex, which catalyzes the first step in fatty acid biosynthesis. Four suppressor mutations mapped to this operon. Two alleles were identified as mutations in the accC gene, the third allele was identified as a mutation in the accB gene, and the fourth allele was shown to be an insertion of an IS1 transposable element in the promoter region of the operon, resulting in reduced transcription. The suppressor mutations caused a decrease in the rate of phospholipid biosynthesis, restoring the balance between the biosynthesis of phospholipids and growth rate, thus enabling *htrB* bacteria to grow at high temperatures.

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3/3,AB/20 (Item 20 from file: 440)  
 DIALOG(R)File 440:Current Contents Search(R)  
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03470266 GENUINE ARTICLE#: HE459 NUMBER OF REFERENCES: 38

TITLE: ISOLATION AND CHARACTERIZATION OF THE ESCHERICHIA-COLI MSBB  
 GENE, A MULTICOPY SUPPRESSOR OF NULL MUTATIONS IN THE  
 HIGH-TEMPERATURE REQUIREMENT GENE HTRB

AUTHOR(S): KAROW M; GEORGOPOULOS C

CORPORATE SOURCE: UNIV UTAH, SCH MED, DEPT CELLULAR VIRAL & MOLEC BIOL/SALT  
 LAKE CITY//UT/84132 (Reprint)

PUBLICATION: JOURNAL OF BACTERIOLOGY, 1992, V174, N3 (FEB), P702-710

LANGUAGE: ENGLISH DOCUMENT TYPE: ARTICLE

ABSTRACT: Previous work established that the *htrB* gene of *Escherichia coli* is required for growth in rich media at temperatures above 32.5-degrees-C but not at lower temperatures. In an effort to determine the functional role of the *htrB* gene product, we have isolated a multicopy suppressor of *htrB*, called *msbB*. The *msbB* gene has been mapped to 40.5 min on the *E. coli* genetic map, in a 12- to 15-kb gap of the genomic library made by Kohara et al. (Y. Kohara, K. Akiyama, and K. Isono, Cell 50:495-508, 1987). Mapping data show that the order of genes in the region is *eda-edd-zwf-pykA-msbB*. The *msbB* gene codes for a protein of 37,410 Da whose amino acid sequences is similar to that of *HtrB* and, like *HtrB*, the protein is very basic in nature. The similarity of the *HtrB* and *MsbB* proteins could indicate that they play functionally similar roles. Mutational analysis of *msbB* shows that the gene is not essential for *E. coli* growth; however, the *htrB msbB* double mutant exhibits a unique morphological phenotype at 30-degrees-C not seen with either of the single mutants. Analysis of both *msbB* and *htrB* mutants shows that these bacteria are resistant to four times more deoxycholate than wild-type bacteria but not to other hydrophobic substances. The addition of quaternary ammonium compounds rescues the temperature-sensitive phenotype of

Searcher : Shears 308-4994

**htrB** bacteria, and this rescue is abolished by the simultaneous addition of Mg<sup>2+</sup> or Ca<sup>2+</sup>. These results suggest that **MsbB** and **HtrB** play an important role in outer membrane structure and/or function.

3/3,AB/21 (Item 21 from file: 440)  
 DIALOG(R)File 440:Current Contents Search(R)  
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03122489 GENUINE ARTICLE#: GG051 NUMBER OF REFERENCES: 33  
 TITLE: SEQUENCING, **MUTATIONAL** ANALYSIS, AND TRANSCRIPTIONAL  
 REGULATION OF THE *ESCHERICHIA-COLI* **HTRB** GENE  
 AUTHOR(S): KAROW M; GEORGOPOULOS C  
 CORPORATE SOURCE: UNIV UTAH, SCH MED, DEPT CELLULAR VIRAL & MOLEC BIOL/SALT  
 LAKE CITY//UT/84132 (Reprint)  
 PUBLICATION: MOLECULAR MICROBIOLOGY, 1991, V5, N9 (SEP), P2285-2292  
 LANGUAGE: ENGLISH DOCUMENT TYPE: ARTICLE  
 ABSTRACT: The *Escherichia coli* **htrB** gene was originally  
 discovered because its insertional inactivation led to an exquisitely  
 temperature-sensitive phenotype in rich media, i.e. the ability to form  
 colonies at temperatures below 32-degrees-C, but not above  
 33-degrees-C. The **htrB** gene has been sequenced. It can  
 potentially code for two proteins, with M(r) values of 35407 Da and  
 8669 Da, that are encoded by overlapping, divergent open reading  
 frames. Our data are consistent with the 35407 Da protein being  
**HtrB**. Northern blot analysis clearly shows that the  
 monocistronic **htrB** message is not under heat-shock regulation.  
 We have also sequenced the flanking DNA and have discovered a new gene,  
 designated orf39.9, located immediately adjacent to **htrB**, but  
 divergently transcribed.

3/3,AB/22 (Item 22 from file: 440)  
 DIALOG(R)File 440:Current Contents Search(R)  
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02822742 GENUINE ARTICLE#: FM035 NUMBER OF REFERENCES: 30  
 TITLE: COMPLEX PHENOTYPES OF NULL **MUTATIONS** IN THE **HTR** GENES, WHOSE  
 PRODUCTS ARE ESSENTIAL FOR *ESCHERICHIA-COLI* GROWTH AT ELEVATED  
 TEMPERATURES  
 AUTHOR(S): KAROW M; RAINA S; GEORGOPOULOS C (Reprint); FAYET O  
 CORPORATE SOURCE: UNIV UTAH, MED CTR, DEPT CELLULAR VIRAL & MOLEC BIOL/SALT  
 LAKE CITY//UT/84132 (Reprint); UNIV UTAH, MED CTR, DEPT CELLULAR VIRAL &  
 MOLEC BIOL/SALT LAKE CITY//UT/84132; CNRS, CTR RECH BIOCHIM & GENET  
 CELLULAIRES/F-31062 TOULOUSE//FRANCE/  
 PUBLICATION: RESEARCH IN MICROBIOLOGY, 1991, V142, N2-3 (FEB-APR), P289-294  
 LANGUAGE: ENGLISH DOCUMENT TYPE: ARTICLE  
 ABSTRACT: Transposon insertion, followed by screening, has allowed the  
 Searcher : Shears 308-4994

identification of a set of genes, called *htr*, whose products are required for *Escherichia coli* growth at elevated temperatures. The *htrB* gene has been shown to map at 23.5 min on the *E. coli* genetic map. It codes for a very basic, hydrophobic, 35,000-Mr polypeptide, possessing a putative membrane-spanning domain. At the non-permissive temperature, *htrB* mutant bacteria stop dividing, followed by the formation of bulges and eventual lysis. The *htrC* gene maps at 90 min, is under sigma-32 regulation and codes for a 21, 130-Mr polypeptide. At 43-degrees-C, *htrC* mutant bacteria gradually lyse, whereas at intermediate temperatures they filament extensively. Finally, the *htrM* gene maps at 81 min, is under sigma-32 regulation and codes for a 35,000-Mr polypeptide. The *HtrM* null phenotype included inability to grow above 42-degrees-C, extreme mucoidness and sensitivity to bile salts, even at the permissive temperatures. The *htrM* gene is identical to the *rfaD* gene, whose product is required for the biosynthesis of the lipopolysaccharide precursor ADP-L-glycero-D-mannoheptose (Pegues et al., J. Bact., 1990, 172, 4652-4660).

3/3,AB/23 (Item 23 from file: 440)  
 DIALOG(R)File 440:Current Contents Search(R)  
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02530639 GENUINE ARTICLE#: ET446 NUMBER OF REFERENCES: 62

TITLE: ISOLATION AND CHARACTERIZATION OF THE ESCHERICHIA-COLI

*HTRB* GENE, WHOSE PRODUCT IS ESSENTIAL FOR BACTERIAL VIABILITY  
 ABOVE 33-DEGREES-C IN RICH MEDIA

AUTHOR(S): KAROW M; FAYET O; CEGIELSKA A; ZIEGELHOFFER T; GEORGOPOULOS C  
 CORPORATE SOURCE: UNIV UTAH,SCH MED,DEPT CELLULAR VIRAL & MOLEC BIOL/SALT  
 LAKE CITY//UT/84132 (Reprint); CNRS,CTR RECH BIOCHIM & GENET  
 CELLULAIRES/F-31062 TOULOUSE//FRANCE/

PUBLICATION: JOURNAL OF BACTERIOLOGY, 1991, V173, N2 (JAN), P741-750

LANGUAGE: ENGLISH DOCUMENT TYPE: ARTICLE

ABSTRACT: We have identified and studied the *htrB* gene of *Escherichia coli*. Insertional inactivation of the *htrB* gene leads to bacterial death at temperatures above 33-degrees-C. The mutant bacterial phenotype at nonpermissive temperatures includes an arrest of cell division followed by the formation of bulges or filaments. The *htrB+* gene has been cloned by complementation and shown to reside at 23.4 min on the *E. coli* genetic map, the relative order of the neighboring loci being *mboA-htrB-pyrC*. The *htrB* gene is transcribed in a counterclockwise fashion, relative to the *E. coli* genetic map, and its product has been identified as a membrane-associated protein of 35,000 Da. Growth experiments in minimal media indicate that the *HtrB* function becomes dispensable at low growth rates.

09/077572

3/3,AB/24 (Item 1 from file: 144)  
DIALOG(R)File 144:Pascal  
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11999709 PASCAL No.: 95-0187031  
Molecular cloning and characterization of the nontypeable  
*Haemophilus influenzae* 2019 rfaE gene required for lipopolysaccharide  
biosynthesis

NA-GYONG LEE; SUNSHINE M G; APICELLA M A  
Univ. Iowa, dep. microbiology, Iowa City IA 52242, USA  
Journal: Infection and immunity, 1995, 63 (3) 818-824  
Language: English

The lipooligosaccharide (LOS) of nontypeable *Haemophilus influenzae* (NTHi) is an important factor in pathogenesis and virulence. In an attempt to elucidate the genes involved in LOS biosynthesis and virulence. In an attempt to elucidate the genes involved in LOS biosynthesis, we have cloned the rfaE gene from NTHi 2019 by complementing a *Salmonella typhimurium* rfaE mutant strain with an NTHi 2019 plasmid library. The rfaE mutant synthesizes lipopolysaccharide (LPS) lacking heptose, and the rfaE gene is postulated to be involved in ADP-heptose synthesis. Retransformation with the plasmid containing 4 kb of NTHi DNA isolated from a reconstituted mutant into rfaE mutants gave wild-type LPS phenotypes. Sodium dodecyl sulfate-polyacrylamide gel electrophoresis analysis confirmed the conversion of the rfaE mutant LPS to a wild-type LPS phenotype. Sequence analysis of a 2.4-kb BglIII fragment revealed two open reading frames. One open reading frame encodes the RfaE protein with a molecular weight of 37.6 kDa, which was confirmed by in vitro transcription and translation, and the other encodes a polypeptide highly homologous to the *Escherichia coli* HtrB protein. These two genes are transcribed from the same promoter region into opposite directions. Primer extension analysis of the rfaE gene revealed a single transcription start site at 37 bp upstream of the predicted translation start site. The upstream promoter region contained a sequence (TA AAAT) homologous to the -10 region of the bacterial SUP 7 SUP 0 -dependent promoters at an appropriate distance (7 bp), but no sequence resembling the consensus sequence of the -35 region was found. These studies demonstrate the ability to use complementation of defined LPS defects in members of the family Enterobacteriaceae to identify LOS synthesis genes in NTHi

3/3,AB/25 (Item 1 from file: 98)  
DIALOG(R)File 98:General Sci Abs/Full-Text  
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03051075 H.W. WILSON RECORD NUMBER: BGSI95051075  
How *Salmonella* survive against the odds.

Foster, John W

Spector, Michael P

Annual Review of Microbiology (Annu Rev Microbiol) v. 49 ('95) p. 145-74

DOCUMENT TYPE: Feature Article

Searcher : Shears 308-4994

SPECIAL FEATURES: bibl il ISSN: 0066-4227

LANGUAGE: English

COUNTRY OF PUBLICATION: United States

WORD COUNT: 14614

**ABSTRACT:** The enteric pathogen *Salmonella typhimurium* faces daunting odds during its voyages in the natural environment and through an infected host. It must manage stresses ranging from feast to famine, acid to base, and high to low osmolarity, among others, as well as counter various types of oxidative stress and a variety of antimicrobial peptides. The defenses used to survive these encounters can be specific or can provide cross protection to a variety of hostile conditions. Once inside a host, *Salmonella* spp. escape the extracellular environment and thus humoral immunity by invading professional and nonprofessional phagocytes in which a new set of challenges await. Some of these stresses are similar to those encountered in the natural environment (e.g. acid, starvation) but the bacterial response is complicated by the simultaneous occurrence of multiple stresses. *S. typhimurium* appears to sense various in vivo cues and responds by seducing the host signal-transduction pathways that are required to phagocytize the bacterial cell. The pathogen then calls upon components of its stress-response arsenal to survive the intracellular environment. These survival strategies enable the organism to persist in nature, where conditions are usually suboptimal and equip the bacterium with pathogenic properties that, if successful, will provide it with a very rich and stress-free growth environment, a dead host. Reprinted by permission of the publisher.

3/3,AB/26 (Item 1 from file: 35)

DIALOG(R) File 35:Dissertation Abstracts Online

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01252959 AAD9234745

MOLECULAR GENETICS OF THE *ESCHERICHIA COLI* *htrB* GENE

Author: KAROW, MARGARET LEE

Degree: PH.D.

Year: 1992

Corporate Source/Institution: THE UNIVERSITY OF UTAH (0240)

Source: VOLUME 53/07-B OF DISSERTATION ABSTRACTS INTERNATIONAL.

PAGE 3313. 182 PAGES

The *Escherichia coli htrB* gene was identified during an insertional mutagenesis screen for new heat shock genes. *HtrB* is essential for viability in rich media only at temperatures above 32.5°C, a phenotype related to bacterial growth rate, since *htrB* bacteria are viable at high temperatures in minimal medium. Despite its unique temperature-sensitive phenotype, the *htrB* gene is not under heat shock regulation. When grown at nonpermissive temperatures, *htrB* bacteria exhibit density-dependent morphological alterations,

Searcher : Shears 308-4994



including the formation of bulges and filaments. The lipopolysaccharide layer of the outer membrane may be altered in *htrB* bacteria, as indicated by the ability of cationic detergents to reverse the lethal phenotype and by the increased resistance of *htrB* bacteria to deoxycholate.

Four spontaneously arising mutations that suppress the *HtrB* temperature-sensitive phenotype were mapped to the *accBC* operon, encoding two of the subunits of acetyl-CoA carboxylase, which catalyzes the first step in fatty acid biosynthesis. Biochemical analysis indicates that *htrB* mutant bacteria overproduce phospholipids at nonpermissive temperatures, a phenotype closely correlated with loss in viability. The *accBC* mutations most likely suppress the lethal phenotype of *htrB* by lowering the rate of fatty acid biosynthesis, thus inhibiting the phospholipid overproduction.

Two new genes were also identified in this study as multicopy suppressors of *htrB*. The protein encoded by the *msbA* suppressor is related to the ATP-dependent translocator family of proteins involved in the export of molecules out of cells. The *msbA* gene is a unique member of this family because it is essential for bacterial viability. The *orfE* gene, which is coexpressed with *msbA*, is also essential. The protein encoded by the *msbB* suppressor gene appears to play a similar, if not redundant role to *HtrB*, because *MsbB* and *HtrB* have similar amino acid sequences and structural feature. Furthermore, *htrB* *msbB* double mutant bacteria exhibit both morphological alterations and growth defects at 30°C, phenotypes that are not exhibited by either of the single mutants. Although bacteria with *msbB* null mutations are viable, they also exhibit an increased resistance to deoxycholate, indicating that like *HtrB*, *MsbB* may play a role in outer membrane function.

? ds

Set	Items	Description
S4	338	AU=(APICELLA, M? OR APICELLA M?)
S5	87	AU=(SUNSHINE, M? OR SUNSHINE M?)
S6	2431	AU=(LEE, N? OR LEE N?)
S7	28	AU=(ARUMUGHAM R? OR ARUMUGHAM, R?)
S8	931	AU=(GIBSON, B? OR GIBSON B?)
S9	0	S4 AND S5 AND S6 AND S7 AND S8
S10	74	S4 AND (S5 OR S6 OR S7 OR S8)
S11	23	S5 AND (S6 OR S7 OR S8)
S12	10	S6 AND (S7 OR S8)
S13	0	S7 AND S8
S14	3708	S4 OR S5 OR S6 OR S7 OR S8
S15	25	(S10 OR S14) AND S1
S16	33	S11 OR S12 OR S15
S18	26	S16 NOT S3

- Author (s)

S19 7 RD (unique items)  
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19/3,AB/1 (Item 1 from file: 440)  
 DIALOG(R)File 440:Current Contents Search(R)  
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08310828 GENUINE ARTICLE#: WQ298 NUMBER OF REFERENCES: 26

TITLE: Identification of the ADP-L-glycero-D-manno-heptose-6-epimerase  
 (rfaD) and heptosyltransferase II (rfaF) biosynthesis genes from  
 nontypeable *Haemophilus influenzae* 2019

AUTHOR(S): Nichols WA; Gibson BW; Melaugh W; Lee NG;  
 Sunshine M; Apicella MA (REPRINT)

CORPORATE SOURCE: UNIV IOWA, COLL MED, DEPT MICROBIOL, BSB 3-403, 51 NEWTON  
 RD/IOWA CITY//IA/52242 (REPRINT); UNIV IOWA, COLL MED, DEPT  
 MICROBIOL/IOWA CITY//IA/52242; UNIV CALIF SAN FRANCISCO, SCH PHARM, DEPT  
 PHARMACEUT CHEM/SAN FRANCISCO//CA/94143; UNIV SAN FRANCISCO, DEPT  
 CHEM/SAN FRANCISCO//CA/94117

PUBLICATION TYPE: JOURNAL

PUBLICATION: INFECTION AND IMMUNITY, 1997, V65, N4 (APR), P1377-1386

PUBLISHER: AMER SOC MICROBIOLOGY, 1325 MASSACHUSETTS AVENUE, NW,  
 WASHINGTON, DC 20005-4171

ISSN: 0019-9567

LANGUAGE: English DOCUMENT TYPE: ARTICLE

ABSTRACT: *Haemophilus influenzae* is an important human pathogen, The  
 lipooligosaccharide (LOS) of *H. influenzae* has been implicated as a  
 virulence determinant, To better understand the assembly of LOS in  
 nontypeable *H. influenzae* (NtHi), we have cloned and characterized the  
 rfaD and rfaF genes of NtHi 2019, which encode the  
 ADP-L-glycero-D-manno-heptose-6-epimerase and heptosyltransferase II  
 enzymes, respectively, This cloning was accomplished by the  
 complementation of *Salmonella typhimurium* lipopolysaccharide (LPS)  
 biosynthesis gene mutants, These deep rough mutants are novobiocin  
 susceptible until complemented with the appropriate gene, In this  
 manner, we are able to use novobiocin resistance to select for specific  
 NtHi LOS inner core biosynthesis genes, Such a screening system yielded  
 a plasmid with a 4,8-kb insert, This plasmid was able to complement  
 both rfaD and rfaF mutants of *S. typhimurium*. The LPS of these  
 complemented strains appeared identical to the wild-type *Salmonella*  
 LPS, The genes encoding the rfaD and rfaF genes from NtHi 2019 were  
 sequenced and found to be similar to the analogous genes from *S.*  
*typhimurium* and *Escherichia coli*, The rfaD gene encodes a polypeptide  
 of 35 kDa and the rfaF encodes a protein of 39 kDa, as demonstrated by  
 in vitro transcription-translation studies. Isogenic mutants which  
 demonstrated truncated LOS consistent with inner core biosynthesis  
 mutants were constructed in the NtHi strain 2019, Primer extension  
 analysis demonstrated the presence of a strong promoter upstream of  
 rfaD but suggested only a very weak promoter upstream of rfaF,

Searcher : Shears 308-4994

Complementation studies, however, suggest that the *rfaF* gene does have an independent promoter, Mass spectrometric analysis shows that the LOS molecules expressed by *H. influenzae* *rfaD* and *rfaF* mutant strains have identical molecular masses. Additional studies verified that in the *rfaD* mutant strain, D-glycero-D-mannoheptose is added to the LOS molecule in place of the usual L-glycero-D-manno-heptose. Finally, the genetic organizations of the inner core biosynthesis genes of *S. typhimurium*, *E. coli*, and several strains of *H. influenzae* were examined, and substantial differences were uncovered.

ISSN: 0019-9567

19/3,AB/2 (Item 1 from file: 71)  
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00736156 97240526

Study of the role of the *htrB* gene in *Salmonella typhimurium*  
 virulence

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Journal: Infection and Immunity, 65/11 (4778-4783), 1997, United States

PUBLICATION DATE: 19970000

CODEN: INFIB

ISSN: 0019-9567

DOCUMENT TYPE: Article

LANGUAGES: English SUMMARY LANGUAGES: English

NO. OF REFERENCES: 70

We have undertaken a study to investigate the contribution of the *htrB* gene to the virulence of pathogenic *Salmonella typhimurium*. An *htrB*::mini-Tn10 mutation from *Escherichia coli* was transferred by transduction to the mouse-virulent strain *S. typhimurium* SL1344 to create an *htrB* mutant. The *htrB* mutant was inoculated into mice and found to be severely limited in its ability to colonize organs of the lymphatic system and to cause systemic disease in mice. A variety of experiments were performed to determine the possible reasons for this loss of virulence. Serum killing assays revealed that the *S. typhimurium htrB* mutant was as resistant to killing by complement as the wild-type strain. However, macrophage survival assays revealed that the *htrB* mutant was more sensitive to the intracellular environment of murine macrophages than the wild-type strain. In addition, the bioactivity of the lipopolysaccharide (LPS) of the *htrB* mutant was reduced compared to that of the LPS from the parent strain as measured by both a *Limulus amoebocyte* lysate endotoxin quantitation assay and a tumor necrosis factor alpha bioassay. These

Searcher : Shears 308-4994

results indicate that the **htrB** gene plays a role in the virulence of *S. typhimurium*.

19/3,AB/3 (Item 2 from file: 71)  
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00736108 97240478

Evaluation of the virulence of nontypeable *Haemophilus influenzae* lipooligosaccharide **htrB** and *rfaD* mutants in the chinchilla model of otitis media

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Journal: Infection and Immunity, 65/11 (4431-4435), 1997, United States

PUBLICATION DATE: 19970000

CODEN: INFIB

ISSN: 0019-9567

DOCUMENT TYPE: Article

LANGUAGES: English SUMMARY LANGUAGES: English

NO. OF REFERENCES: 30

Considerable evidence has implicated nontypeable *Haemophilus influenzae* (NTHi) lipooligosaccharide (LOS) in the pathogenesis of otitis media (OM); however, its exact role has not been conclusively established. Recently, two NTHi LOS-deficient mutants have been created and described. Strain 2019-DK1, an *rfaD* gene mutant, expresses a truncated LOS consisting of only three deoxy-D-manno-octulosonic acid residues, a single heptose, and lipid A. Strain 2019-B29, an isogenic **htrB** mutant, possesses an altered oligosaccharide core and an altered lipid A. Each strain's ability to colonize the nasopharynx and to induce OM subsequent to transbullar inoculation was evaluated in the chinchilla model. Nasopharyngeal colonization data indicate that the parent strain and both mutants are able to colonize the nasopharynx and exhibit comparable clearance kinetics. Compared with the parent and each other, however, the mutants demonstrated marked differences in virulence regarding their relative abilities to induce OM and persist in the middle ear post-transbullar inoculation. Strain B29 required a 3-log-greater dose to induce OM than the parent strain and did not exhibit evidence of sustained multiplication but persisted for the same duration as the parent. Conversely, strain-DK1, even when inoculated at a dose 4 logs greater than the parent dose, was eliminated from the middle ear 72 h after challenge. A comparison of the relative pathogenicities of these isolates provides the opportunity to address fundamental questions regarding the contribution of LOS to pathogenesis issues at the molecular level. Specifically, the impact of these LOS gene disruptions on OM pathogenesis can be defined and may thus

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provide potential new targets for future protection and intervention strategies.

19/3,AB/4 (Item 3 from file: 71)  
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00699175 97202783

Mutation of the *htrB* gene in a virulent *Salmonella typhimurium* strain by intergeneric transduction: Strain construction and phenotypic characterization

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EMAIL: michael-apicella@uiowa.edu

Journal: Journal of Bacteriology, 179/17 (5521-5533), 1997, United States

PUBLICATION DATE: 19970000

CODEN: JOBAA

ISSN: 0021-9193

DOCUMENT TYPE: Article

LANGUAGES: English

SUMMARY LANGUAGES: English

NO. OF REFERENCES: 36

The *htrB* gene product of *Haemophilus influenzae* contributes to the toxicity of the lipooligosaccharide. The *htrB* gene encodes a 2-keto-3-deoxyoctulosonic acid-dependent acyltransferase which is responsible for myristic acid substitutions at the hydroxy moiety of lipid A beta-hydroxymyristic acid. Mass spectroscopic analysis has demonstrated that lipid A from an *H. influenzae htrB* mutant is predominantly tetraacyl and similar in structure to lipid IV(A), which has been shown to be nontoxic in animal models. We sought to construct a *Salmonella typhimurium htrB* mutant in order to investigate the contribution of *htrB* to virulence in a well-defined murine typhoid model of animal pathogenesis. To this end, an *rsup* - *msup* + *gale mutS recD* strain of *S. typhimurium* was constructed (MGS-7) and used in inter- and intrastrain transduction experiments with both coliphage P1 and *Salmonella* phage P22. The *Escherichia coli htrB* gene containing a mini-Tn10 insertion was transduced from *E. coli* MLK217 into *S. typhimurium* MGS-7 via phage P1 and subsequently via phage P22 into the virulent *Salmonella* strain SL1344. All *S. typhimurium* transductants showed phenotypes similar to those described for the *E. coli htrB* mutant. Mass spectrometric analysis of the crude lipid A fraction from the lipopolysaccharide of the *S. typhimurium htrB* mutant strain showed that for the dominant hexaacyl form, a lauric acid moiety was lost at one position on the lipid A and a palmitic acid moiety was added at another position; for the less abundant heptaacyl species, the lauric acid was replaced with palmitoleic acid.

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19/3,AB/5 (Item 4 from file: 71)  
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00680306 97187075

**htrB** of *Haemophilus influenzae*: Determination of biochemical

activity and effects on virulence and lipooligosaccharide toxicity

Nichols W.A.; Raetz C.R.H.; Clementz T.; Smith A.L.; Hanson J.A.; Ketterer M.R.; **Sunshine M.**; **Apicella M.A.**

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Journal: Journal of Endotoxin Research, 4/3 (163-172), 1997, United Kingdom

PUBLICATION DATE: 19970000

CODEN: JENRE

ISSN: 0968-0519

DOCUMENT TYPE: Article

LANGUAGES: English SUMMARY LANGUAGES: English

NO. OF REFERENCES: 28

The **htrB** mutant of *Haemophilus influenzae* (strain B29) has been shown to lack secondary (nonhydroxylated) acyl groups in its lipid A. We have determined through in vitro biochemical assays that the **HtrB** protein acts as a specific acyltransferase in the late stages of lipid A biosynthesis and that the preferred acyl group donor is myristoyl-acyl carrier protein. Under the conditions employed, the *Escherichia coli* precursor, Kdoinf 2-lipid IV(A), functions as a myristate acceptor. Introduction of the *Haemophilus htrB* gene into an *E. coli* mutant lacking **htrB** complements the biochemical and physiological defects associated with the *E. coli htrB* mutation. Tumor necrosis factor alpha (TNFalpha) assays using murine and human macrophage cells indicated that nontypeable *H. influenzae* (NtHi) strain 2019 and *H. influenzae* type b strain A2 elicit levels of expression of TNFalpha that are 30-40 times greater than levels induced by the isogenic **htrB** mutants (B29 and A2B29). Studies using cell-free LOS indicated that the LOS from wild type strain 2019 elicits levels of TNFalpha expression that are 6-8-fold higher than those of B29. In situ hybridization studies of a primary human bronchial epithelial cell line demonstrated a greater increase of TNFalpha message produced in the presence of 2019 LOS than in the presence of B29 LOS. TNFalpha levels of the cell supernatant of cells stimulated with 2019 LOS were found to be 7-8-fold higher than levels in B29 stimulated supernatants. Using the *Limulus* amoebocyte lysate for assessment of endotoxic activity, we found that wild type LOS was 8-fold higher in endotoxic activity compared with the mutant LOS. In virulence assays using intraperitoneal inoculation of infant rats, the **htrB** isogenic strain caused bacteremia at 50% the frequency of the wild type

Searcher : Shears 308-4994

strain. In intranasal inoculation studies, the *htrB* mutant strain was unable to cause bacteremia whereas the wild type b parent produced bacteremia in 40-60% of the animals. These findings suggest that the *htrB* gene of *H. influenzae* is important for virulence and that host TNFalpha expression is attenuated in response to *htrB* mutant strains.

19/3,AB/6 (Item 5 from file: 71)  
 DIALOG(R) File 71:ELSEVIER BIOBASE  
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00323784 96007283

Mutation of the *htrB* locus of *Haemophilus influenzae* nontypable strain 2019 is associated with modifications of lipid A and phosphorylation of the lipo-oligosaccharide

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Journal: Journal of Biological Chemistry, 270/45 (27151-27159), 1995,  
 United States

PUBLICATION DATE: 19950000

CODEN: JBCHA

ISSN: 0021-9258

DOCUMENT TYPE: Article

LANGUAGES: English SUMMARY LANGUAGES: English

The *HtrB* protein was first identified in *Escherichia coli* as a protein required for cell viability at high temperature, but its expression was not regulated by temperature. We isolated an *htrB* homologue from non-typable *Haemophilus influenzae* strain (NTHi) 2019, which was able to functionally complement the *E. coli htrB* mutation. The promoter for the NTHi 2019 *htrB* gene overlaps the promoter for the *rfaE* gene, and the two genes are divergently transcribed. The deduced amine acid sequence of NTHi 2019 *HtrB* had 56% homology to *E. coli HtrB*. In vitro transcription-translation analysis confirmed production of a protein with an apparent molecular mass of 32-33 kDa. Primer extension analysis revealed that *htrE* was transcribed from sigma<sup>70</sup>-dependent consensus promoter and its expression was not affected by temperature. The expression of *htrB* and *rfaE* was 2.5-4 times higher in the NTHi *htrB* mutant B29 than in the parental strain. In order to study the function of the *HtrB* protein in *Haemophilus*, we generated two isogenic *htrB* mutants by shuttle mutagenesis using a mini-Tn3. The *htrB* mutants initially showed temperature sensitivity, but they lost the sensitivity after a few passages at 30 degreeC and were able to grow at 37 degreeC. They also showed hypersensitivity to deoxycholate and kanamycin, which persisted on passage. SDS-polyacrylamide gel electrophoresis analysis revealed that the lipo- oligosaccharide (LOS)

Searcher : Shears 308-4994

isolated from these mutants migrated faster than the wild type LOS and its color changed from black to brown as has been described for *E. coli* **htrB** mutants. Immunoblotting analysis also showed that the LOS from the **htrB** mutants lost reactivity to a monoclonal antibody, 6E4, which binds to the wild type NTHi 2019 LOS. Electrospray ionization-mass spectrometry analysis of the O-deacylated LOS oligosaccharide indicated a modification of the core structure characterized in part by a net loss in phosphoethanolamine. Mass spectrometric analysis of the lipid A of the **htrB** mutant indicated a loss of one r both myristic acid substitutions. These data suggest that **HtrB** is a multifunctional protein and may play a controlling role in regulating cell responses to various environmental changes.

19/3,AB/7 (Item 6 from file: 71)  
 DIALOG(R)File 71:ELSEVIER BIOBASE  
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00248954 95047178

Molecular cloning and characterization of the nontypeable **Haemophilus influenzae** 2019 **rfaE** gene required for lipopolysaccharide biosynthesis  
 Lee N.-G.; Sunshine M.G.; Apicella M.A.

ADDRESS: M.A. Apicella, Department of Microbiology, Bowen Science Building,  
 University of Iowa, 3-401, 51 Newton Road, Iowa City, IA 52242,  
 United States

Journal: Infection and Immunity, 63/3 (818-824), 1995, United States

PUBLICATION DATE: 19950000

CODEN: INFIB

ISSN: 0019-9567

DOCUMENT TYPE: Article

LANGUAGES: English SUMMARY LANGUAGES: English

The lipooligosaccharide (LOS) of nontypeable **Haemophilus influenzae** (NTHi) is an important factor in pathogenesis and virulence. In an attempt to elucidate the genes involved in LOS biosynthesis, we have cloned the **rfaE** gene from NTHi 2019 by complementing a **Salmonella typhimurium** **rfaE** mutant strain with an NTHi 2019 plasmid library. The **rfaE** mutant synthesizes lipopolysaccharide (LPS) lacking heptose, and the **rfaE** gene is postulated to be involved in ADP-heptose synthesis. Retransformation with the plasmid containing 4 kb of NTHi DNA isolated from a reconstituted mutant into **rfaE** mutants gave wild-type LPS phenotypes. Sodium dodecyl sulfate-polyacrylamide gel electrophoresis analysis confirmed the conversion of the **rfaE** mutant LPS to a wild-type LPS phenotype. Sequence analysis of a 2.4-kb BglII fragment revealed two open reading frames. One open reading frame encodes the **RfaE** protein with a molecular weight of 37.6 kDa, which was confirmed by in vitro transcription and translation, and the other encodes a polypeptide highly homologous to the *Escherichia coli* **HtrB** protein. These two genes are transcribed from the same promoter region into opposite directions. Primer extension analysis of the **rfaE** gene

Searcher : Shears 308-4994



09/077572

revealed a single transcription start site at 37 bp upstream of the predicted translation start site. The upstream promoter region contained a sequence (TAAAT) homologous to the -10 region of the bacterial *delta*<sup>sup</sup> 0-dependent promoters at an appropriate distance (7 bp), but no sequence resembling the consensus sequence of the -35 region was found. These studies demonstrate the ability to use complementation of defined LPS defects in members of the family Enterobacteriaceae to identify LOS synthesis genes in NTHi.

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01mar99 11:16:37 User219783 Session.D1447.4

09/077572

(FILE 'USPAT' ENTERED AT 10:44:05 ON 01 MAR 1999)

Key Terms

L1 26 SEA HTRB OR HTR B  
L2 1 SEA L1(5A) (MUTAT? OR MUTANT# OR MUTAGEN? OR POLYMORPH? OR  
POLY MORPHI###)  
L3 10 SEA L1 AND (COLI OR GRAM(W) (NEGATIVE OR NEG) OR SALMONELL?  
OR HAEMOPHIL? OR HEMOPHIL?)  
L4 10 SEA L2 OR L3

US PAT NO: 5,846,723 [IMAGE AVAILABLE] L4: 1 of 10  
TITLE: Methods for detecting the RNA component of telomerase  
DATE ISSUED: Dec. 8, 1998  
INVENTOR: Nam Woo Kim, San Jose, CA  
Fred Wu, San Carlos, CA  
James T. Kealey, San Anselmo, CA  
Ronald Pruzan, Palo Alto, CA  
Scott L. Weinrich, Redwood City, CA  
SEARCH-FLD: 435/6, 91.2, 91.3; 536/23.1, 24.3-24.33

## ABSTRACT:

Methods of detecting the RNA component of telomerase, diagnosing cancer, and determining its prognosis using polynucleotides that hybridize to the RNA component of mammalian telomerase in a sample.

US PAT NO: 5,824,538 [IMAGE AVAILABLE] L4: 2 of 10  
TITLE: Shigella vector for delivering DNA to a mammalian cell  
DATE ISSUED: Oct. 20, 1998  
INVENTOR: Arthur A. Branstrom, Rockville, MD  
Donata R. Sizemore, Gaithersburg, MD  
Jerald C. Sadoff, Washington, DC  
SEARCH-FLD: 424/234.1, 235.1, 93.2; 435/245, 172.3, 252.1, 252.3, 822,  
172.1

## ABSTRACT:

We describe a bacterial delivery system for the delivery of DNA and antigens into cells. We constructed an attenuated bacterial vector which enters mammalian cells and ruptures delivering functional plasmid DNA, such as a mammalian expression plasmid, and antigens into the cell cytoplasm. This Shigella vector was designed to deliver DNA to colonic surfaces, thus opening the possibility of oral and other mucosal DNA immunization and gene therapy strategies. The attenuated Shigella is also useful as a vaccine for reducing disease symptoms caused by Shigella.

US PAT NO: 5,801,233 [IMAGE AVAILABLE] L4: 3 of 10  
TITLE: Nucleic acid compositions encoding acetyl-coa carboxylase  
and uses therefor  
DATE ISSUED: Sep. 1, 1998  
INVENTOR: Robert Haselkorn, Chicago, IL  
Piotr Gornicki, Chicago, IL  
SEARCH-FLD: 435/172.3, 69.1, 320.1, 252.1, 255.1, 257.1, 325, 410,  
419, 6, 975; 536/23.1, 23.2, 23.4, 23.6, 23.7, 24.3,  
24.32; 935/9, 22, 66, 60, 67, 73  
Searcher : Shears 308-4994

## ABSTRACT:

The present invention provides isolated and purified polynucleotides that encode plant and cyanobacterial polypeptides that participate in the carboxylation of acetyl-CoA. Isolated cyanobacterial and plant polypeptides that catalyze acetyl-CoA carboxylation are also provided. Processes for altering acetyl-CoA carboxylation, increasing herbicide resistance of plants and identifying herbicide resistant variants of acetyl-CoA carboxylase are also provided.

US PAT NO: 5,776,679 [IMAGE AVAILABLE] L4: 4 of 10  
 TITLE: Assays for the DNA component of human telomerase  
 DATE ISSUED: Jul. 7, 1998  
 INVENTOR: Bryant Villeponteau, San Carlos, CA  
 Junli Feng, San Carlos, CA  
 Walter Funk, Union City, CA  
 William H. Andrews, Richmond, CA  
 SEARCH-FLD: 435/6, 91.2, 91.21, 91.51; 436/63, 64; 536/24.31, 23.1,  
 24.33; 935/8, 3, 78

## ABSTRACT:

Nucleic acids comprising the RNA component of a mammalian telomerase are useful as pharmaceutical, therapeutic, and diagnostic reagents.

US PAT NO: 5,599,904 [IMAGE AVAILABLE] L4: 5 of 10  
 TITLE: Chimeric steroid hormone superfamily receptor proteins  
 DATE ISSUED: Feb. 4, 1997  
 INVENTOR: Ronald M. Evans, La Jolla, CA  
 Estelita S. Ong, San Diego, CA  
 Prudimar S. Segui, San Diego, CA  
 Catherine C. Thompson, La Jolla, CA  
 Kazuhiko Umesono, San Diego, CA  
 Vincent Giguere, Etobicoke, Canada  
 SEARCH-FLD: 530/350, 358, 399; 435/69.7, 69.1; 935/36

## ABSTRACT:

A novel retinoic acid receptor is disclosed. The novel receptor is encoded for by cDNA carried on plasmid pHRAR1, which has been deposited with the American Type Culture Collection for patent purposes. Chimeric receptor proteins are also disclosed. The chimera are constructed by exchanging functional domains between the glucocorticoid, the mineralocorticoid, the estrogen-related, the thyroid and the retinoic acid receptors. In addition, a novel method for identifying functional ligands for receptor proteins is disclosed. The method, which takes advantage of the modular structure of the hormone receptors and the idea that the functional domains may be interchangeable, replaces the DNA-binding domain of a putative novel receptor with the DNA-binding domain of a known receptor such as the glucocorticoid receptor. The resulting chimeric construction, when expressed in cells, produces a hybrid receptor whose activation of a ligand-(e.g., glucocorticoid) inducible promoter is dependent on the presence of the new ligand. The

Searcher : Shears 308-4994

novel method is illustrated in part by showing that the ligand for the new receptor protein is the retinoid, retinoic acid.

US PAT NO: 5,571,692 [IMAGE AVAILABLE] L4: 6 of 10  
 TITLE: Retinoic acid receptor .alpha., vectors and cells  
 comprising the same DNA encoding  
 DATE ISSUED: Nov. 5, 1996  
 INVENTOR: Ronald M. Evans, La Jolla, CA  
 Estelita S. Ong, San Diego, CA  
 Prudimar S. Segui, San Diego, CA  
 Catherine C. Thompson, La Jolla, CA  
 Kazuhiko Umesono, San Diego, CA  
 Vincent Giguere, Etobicoke, Canada  
 SEARCH-FLD: 536/23.5, 23.4; 435/240.2, 320.1, 252.3, 254.11, 69.1,  
 69.2

ABSTRACT:

DNA encoding a human retinoic acid receptor alpha protein is disclosed. The sequence of the receptor is encoded by the cDNA insert of plasmid pHRR1, which has been deposited with ATCC. Methods employing chimeric receptors derived from the retinoic acid receptor are illustrated which demonstrate that the ligand for the new receptor is the retinoid, retinoic acid.

US PAT NO: 5,548,063 [IMAGE AVAILABLE] L4: 7 of 10  
 TITLE: Retinoic acid receptor alpha proteins  
 DATE ISSUED: Aug. 20, 1996  
 INVENTOR: Ronald M. Evans, La Jolla, CA  
 Estelita S. Ong, San Diego, CA  
 Prudimar S. Segui, San Diego, CA  
 Catherine C. Thompson, La Jolla, CA  
 Kazuhiko Umesono, San Diego, CA  
 Vincent Giguere, Etobicoke, Canada  
 SEARCH-FLD: 530/350, 324; 435/69.1

ABSTRACT:

A human retinoic acid receptor alpha protein is disclosed. The receptor is encoded by the cDNA insert of plasmid pHRR1, which has been deposited with ATCC. Methods employing chimeric receptors derived from the retinoic acid receptor are illustrated which demonstrate that the ligand for the new receptor is the retinoid, retinoic acid.

US PAT NO: 5,274,077 [IMAGE AVAILABLE] L4: 8 of 10  
 TITLE: Retinoic acid receptor composition  
 DATE ISSUED: Dec. 28, 1993  
 INVENTOR: Ronald M. Evans, La Jolla, CA  
 Estelita S. Ong, San Diego, CA  
 Prudimar S. Segui, San Diego, CA  
 Catherine C. Thompson, La Jolla, CA  
 Kazuhiko Umesono, San Diego, CA  
 Vincent Giguere, Etobicoke, Canada  
 SEARCH-FLD: 530/350, 358; 435/69.7, 252.3

Searcher : Shears 308-4994

## ABSTRACT:

A novel retinoic acid receptor is disclosed. The novel receptor is encoded for by CDNA carried on plasmid pHRAR1, which has been deposited with the American Type Culture Collection for patent purposes. Chimeric receptor proteins are also disclosed. The chimera are constructed by exchanging functional domains between the glucocorticoid, the mineralocorticoid, the estrogen-related, the thyroid and the retinoic acid receptors. In addition, a novel method for identifying functional ligands for receptor proteins is disclosed. The method, which takes advantage of the modular structure of the hormone receptors and the idea that the functional domains may be interchangeable, replaces the DNA-binding domain of a putative novel receptor with the DNA-binding domain of a known receptor such as the glucocorticoid receptor. The resulting chimeric construction, when expressed in cells, produces a hybrid receptor whose activation of a ligand--(e.g., glucocorticoid) inducible promoter is dependent on the presence of the new ligand. The novel method is illustrated in part by showing that the ligand for the new receptor protein is the retinoid, retinoic acid.

US PAT NO: 5,171,671 [IMAGE AVAILABLE] L4: 9 of 10  
 TITLE: Retinoic acid receptor composition  
 DATE ISSUED: Dec. 15, 1992  
 INVENTOR: Ronald M. Evans, La Jolla, CA  
 Estelita S. Ong, San Diego, CA  
 Prudimar S. Segui, San Diego, CA  
 Catherine C. Thompson, La Jolla, CA  
 Kazuhiko Uemsono, San Diego, CA  
 Vincent Giguere, Etobicoke, Canada  
 SEARCH-FLD: 536/27; 435/69.1, 69.7, 252.3, 320.1; 530/350

## ABSTRACT:

A novel retinoic acid receptor is disclosed. The novel receptor is encoded for by cDNA carried on plasmid pHRAR1, which has been deposited with the American Type Culture Collection for patent purposes. Chimeric receptor proteins are also disclosed. The chimera are constructed by exchanging functional domains between the glucocorticoid, the mineralocorticoid, the estrogen-related, the thyroid and the retinoic acid receptors. In addition, a novel method for identifying functional ligands for receptor proteins is disclosed. The method, which takes advantage of the modular structure of the hormone receptors and the idea that the functional domains may be interchangeable, replaces the DNA-binding domain of a putative novel receptor with the DNA-binding domain of a known receptor such as the glucocorticoid receptor. The resulting chimeric construction, when expressed in cells, produces a hybrid receptor whose activation of a ligand-(e.g., glucocorticoid) inducible promoter is dependent on the presence of the new ligand. The novel method is illustrated in part by showing that the ligand for the new receptor protein is the retinoid, retinoic acid.

US PAT NO: 4,981,784 [IMAGE AVAILABLE] L4: 10 of 10  
 Searcher : Shears 308-4994

09/077572

TITLE: Retinoic acid receptor method  
DATE ISSUED: Jan. 1, 1991  
INVENTOR: Ronald M. Evans, La Jolla, CA  
Estelita Ong, San Diego, CA  
Prudimar S. Segui, San Diego, CA  
Catherine C. Thompson, La Jolla, CA  
Kazuhiko Umesono, San Diego, CA  
Vincent Giguere, Etobicoke, Canada  
SEARCH-FLD: 435/6, 29, 41, 172.1, 172.3, 320, 69.1, 69.4, 69.7, 70.1;  
935/6, 11, 9, 13, 23, 27, 70, 76, 111

ABSTRACT:

A novel retinoic acid receptor is disclosed. The novel receptor is encoded for by cDNA carried on plasmid pHRAR1, which has been deposited with the American Type Culture Collection for patent purposes. Chimeric receptor proteins are also disclosed. The chimera are constructed by exchanging functional domains between the glucocorticoid, the mineralocorticoid, the estrogen-related, the thyroid and the retinoic acid receptors. In addition, a novel method for identifying functional ligands for receptor proteins is disclosed. The method, which takes advantage of the modular structure of the hormone receptors and the idea that the functional domains may be interchangeable, replaces the DNA-binding domain of a putative novel receptor with the DNA-binding domain of a known receptor such as the glucocorticoid receptor. The resulting chimeric construction, when expressed in cells, produces a hybrid receptor whose activation of a ligand-(e.g., glucocorticoid) inducible promoter is dependent on the presence of the new ligand. The novel method is illustrated in part by showing that the ligand for the new receptor protein is the retinoid, retinoic acid.

=> d his 15-

(FILE 'USPAT' ENTERED AT 10:44:05 ON 01 MAR 1999)

L5 3 S APICELLA, M?/IN  
L6 1 S SUNSHINE, M?/IN  
L7 250 S LEE, N?/IN  
L8 0 S ARUMUGHAM, R?/AU  
L9 27 S GIBSON, B?/IN  
L10 0 S L5 AND L6 AND L7 AND L9  
L11 0 S L5 AND (L6 OR L7 OR L9)  
L12 0 S L6 AND (L7 OR L9)  
L13 0 S L7 AND L9  
L14 0 S (L5 OR L6 OR L7 OR L9) AND L1

-Author(s)

=> fil hom

FILE 'HOME' ENTERED AT 10:49:35 ON 01 MAR 1999

Searcher : Shears 308-4994

09/077572

FILE 'CAPLUS' ENTERED AT 10:23:40 ON 01 MAR 1999

-key terms

L1 17 SEA ABB=ON PLU=ON (HTRB OR HTR B) (S) (MUTAT? OR  
MUTAGEN? OR MUTANT OR POLYMORPH? OR POLY(W)MORPHI###)  
L2 17 SEA ABB=ON PLU=ON L1 AND (GRAM(W) (NEGATIVE OR NEG) OR  
SALMONELL? OR COLI OR HAEMOPHIL? OR HEMOPHIL?)

L2 ANSWER 1 OF 17 CAPLUS COPYRIGHT 1999 ACS

AN 1999:52080 CAPLUS

TI Lipopolysaccharide from an *Escherichia coli* **htrB**  
**msbB mutant** induces high levels of MIP-1.alpha. and  
MIP-1.beta. secretion without inducing TNF-.alpha. and IL-1.beta.

SO J. Hum. Virol. (1998), 1(4), 251-256

CODEN: JHVIFC; ISSN: 1090-9508

AU Hone, David M.; Powell, Jan; Crowley, Richard W.; Maneval, David;  
Lewis, George K.

PY 1998

AB To identify a lipopolysaccharide (LPS) that retains the capacity to  
induce .beta.-chemokine secretion without the concomitant activation  
of pyrogenic cytokines. LPS was extd. from strain MLK986 (mLPS), an  
htrB1::Tn10, msbB::ocam mutant of *Escherichia coli* that is  
defective for lipid A synthesis, and from wild-type parent *E*  
**coli** strains, W3110 (wtLPS). The capacity of these LPS  
prepns. to induce tumor necrosis factor-.alpha. (TNF-.alpha.),  
interleukin-1.beta. (IL-1.beta.), and macrophage inflammatory  
proteins 1.alpha. (MIP-1.alpha.) and MIP-1.beta. was assessed using  
a human peripheral blood mononuclear cell (PBMC) activation assay.  
Stimulation of PBMCs with mLPS did not induce measurable levels of  
pyrogenic cytokines TNF-.alpha. and IL-1.beta., whereas wtLPS  
induced high levels of these cytokines. Furthermore, mLPS  
antagonized the induction of TNF-.alpha. secretion by wtLPS.  
Nonetheless, mLPS retained a discrete agonist activity that induced  
MIP-1.alpha. and MIP-1.beta. secretion by PBMCs. This latter  
agonist activity appears to be unique to mLPS, since two previously  
documented LPS antagonists, *Rhodobacter sphaeroides* diphosphoryl  
lipid A and synthetic lipid IVA, did not induce MIP-1.alpha. and  
MIP-1.beta. secretion. Furthermore, synthetic lipid IVA was an  
antagonist of MIP-1.alpha. and MIP-1.beta. induction by mLPS. These  
results show that mLPS exhibits a novel bipartite activity, being an  
effective antagonist of TNF-.alpha. induction by wtLPS, while  
paradoxically being an agonist of MIP-1.alpha. and MIP-1.beta.  
secretion.

L2 ANSWER 2 OF 17 CAPLUS COPYRIGHT 1999 ACS

AN 1998:543176 CAPLUS

DN 129:159061

TI **Salmonella** lacking lipid A as a result of **mutation**  
in the **msbB** or **htrB** genes

SO PCT Int. Appl., 39 pp.

CODEN: PIXXD2

Searcher : Shears 308-4994

IN Maskell, Duncan John; Dougan, Gordon

APPLICATION NO. DATE

AI WO 98-GB291 19980130

AU 98-58734 19980130

PATENT NO. KIND DATE APPLICATION NO. DATE

PI WO 9833923 A1 19980806 WO 98-GB291 19980130

W: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ,  
 DE, DK, EE, ES, FI, GB, GE, GM, GW, HU, ID, IL, IS, JP, KE,  
 KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN,  
 MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM,  
 TR, TT, UA, UG, US, UZ, VN, AM, AZ, BY, KG, KZ, MD, RU, TJ,  
 TM

RW: GH, GM, KE, LS, MW, SD, SZ, UG, ZW, AT, BE, CH, DE, DK, ES,  
 FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG,  
 CI, CM, GA, GN, ML, MR, NE, SN, TD, TG

AU 9858734 A1 19980825 AU 98-58734 19980130

PY 1998

1998

AB Two genes, *msbB* and *htrB*, involved in lipid A biosynthesis in *Salmonella* are cloned from *S. typhimurium* and mutations in them are used to create lipid A-deficient strains of *Salmonella*. The lipid A-deficient strains are less toxigenic than wild-type strains and may be used in vaccines. These microorganisms may also be useful as expression hosts. The lipopolysaccharide from these mutants may also be of use as an endotoxin antagonist. Inoculation of mice with an *msbB* mutant of *S. typhimurium* led to .apprx.5% fatalities within 7 days compared with 100% fatality in a group inoculated with the wild-type microorganism. The mutants were less effective at induction of synthesis of tumor necrosis factor .alpha. and interleukin 1.beta. in vitro than were control strains. Double mutant strains (*aro/msbB*) gave 40% protection when used in an oral vaccine compared to 100% protection for an *aroA* single mutant.

L2 ANSWER 3 OF 17 CAPLUS COPYRIGHT 1999 ACS

AN 1998:471252 CAPLUS

DN 129:200426

TI Role of the O-antigen of lipopolysaccharide, and possible roles of growth rate and of NADH:ubiquinone oxidoreductase (nuo) in competitive tomato root-tip colonization by *Pseudomonas fluorescens* WCS365

SO Mol. Plant-Microbe Interact. (1998), 11(8), 763-771

CODEN: MPMIEL; ISSN: 0894-0282

AU Dekkers, Linda C.; Van Der Bij, Arjan J.; Mulders, Ine H. M.;  
 Phoelich, Claartje C.; Wentwoord, Rino A. R.; Glandorf, Deborah C.  
 M.; Wijffelman, Carel A.; Lugtenberg, Ben J. J.

PY 1998

Searcher : Shears 308-4994



AB Colonization-defective, transposon-induced mutants of the efficient root colonizer *Pseudomonas fluorescens* WCS365 were identified with a gnotobiotic system. Most mutants were impaired in known colonization traits, i.e., prototrophy for amino acids, motility, and synthesis of the O-antigen of LPS (lipopolysaccharide). Mutants lacking the O-antigen of LPS were impaired in both colonization and competitive growth, whereas one mutant (PCL1205) with a shorter O-antigen chain was defective only in colonization ability, suggesting a role for the intact O-antigen of LPS in colonization. Eight competitive colonization mutants that were not defective in the above-mentioned traits colonized the tomato root tip well when inoculated alone, but were defective in competitive root colonization of tomato, radish, and wheat, indicating they contained mutations affecting host range. One of these eight mutants (PCL1201) was further characterized and contains a mutation in a gene that shows homol. to the *Escherichia coli* nuo4 gene, which encodes a subunit of one of two known NADH:ubiquinone oxidoreductases. Competition expts. in an oxygen-poor medium between mutant PCL1201 and its parental strain showed a decreased growth rate of mutant PCL1201. The requirement of the nuo4 gene homolog for optimal growth under conditions of oxygen limitation suggests that the root-tip environment is microaerobic. A mutant characterized by a slow growth rate (PCL1216) was analyzed further and contained a mutation in a gene with similarity to the *E. coli* HtrB protein, a lauroyl transferase that functions in lipid A biosynthesis.

L2 ANSWER 4 OF 17 CAPLUS COPYRIGHT 1999 ACS

AN 1998:330380 CAPLUS

DN 129:65382

TI Function of *Escherichia coli* MsbA, an essential ABC family transporter, in lipid A and phospholipid biosynthesis

SO J. Biol. Chem. (1998), 273(20), 12466-12475

CODEN: JBCHA3; ISSN: 0021-9258

AU Zhou, Zhimin; White, Kimberly A.; Polissi, Alessandra; Georgopoulos, Costa; Raetz, Christian R. H.

PY 1998

AB The *Escherichia coli* msbA gene, first identified as a multicopy suppressor of htrB mutations, has been proposed to transport nascent core-lipid A mols. across the inner membrane. MsbA is an essential *E. coli* gene with high sequence similarity to mammalian Mdr proteins and certain types of bacterial ABC transporters. HtrB is required for growth above 32.degree.C and encodes the lauroyltransferase that acts after Kdo addn. during lipid A biosynthesis. By using a quant. new <sup>32</sup>Pi labeling technique, the authors demonstrate that hexa-acylated species of lipid A predominate in the outer membranes of wild type *E. coli* labeled for several generations at 42.degree.C. In contrast, in htrB mutants shifted to

Searcher : Shears 308-4994

42.degree.C for 3 h, tetraacylated lipid A species and glycerophospholipids accumulate in the inner membrane. Extra copies of the cloned msbA gene restore the ability of **htrB** mutants to grow at 42.degree.C, but they do not increase the extent of lipid A acylation. However, a significant fraction of the tetra-acylated lipid A species that accumulate in **htrB** mutants are transported to the outer membrane in the presence of extra copies of msbA. *E. coli* strains in which msbA synthesis is selectively shut off at 42.degree.C accumulate hexa-acylated lipid A and glycerophospholipids in their inner membranes. These results suggest that MsbA plays a role in lipid A and possibly glycerophospholipid transport. The tetra-acylated lipid A precursors that accumulate in **htrB** mutants may not be transported as efficiently by MsbA as are penta- or hexa-acylated lipid A species.

L2 ANSWER 5 OF 17 CAPLUS COPYRIGHT 1999 ACS  
 AN 1997:731002 CAPLUS  
 DN 128:20429  
 TI Study of the role of the htrB gene in *Salmonella*  
*typhimurium* virulence  
 SO Infect. Immun. (1997), 65(11), 4778-4783  
 CODEN: INFIBR; ISSN: 0019-9567  
 AU Jones, Bradely D.; Nichols, Wade A.; Gibson, Bradford W.; Sunshine,  
 Melvin G.; Apicella, Michael A.  
 PY 1997  
 AB We have undertaken a study to investigate the contribution of the  
 htrB gene to the virulence of pathogenic *Salmonella*  
*typhimurium*. An **htrB::mini-Tn10** mutation from  
*Escherichia coli* was transferred by transduction to the  
 mouse-virulent strain *S. typhimurium* SL1344 to create an  
**htrB** mutant. The *S. typhimurium* **htrB**  
 mutant was inoculated into mice and found to be severely  
 limited in its ability to colonize organs of the lymphatic system  
 and to cause systemic disease in mice. A variety of expts. were  
 performed to det. the possible reasons for this loss of virulence.  
 Serum killing assays revealed that the *S. typhimurium* **htrB**  
 mutant was as resistant to killing by complement as the  
 wild-type strain. However, macrophage survival assays revealed that  
 the *S. typhimurium* **htrB** mutant was more  
 sensitive to the intracellular environment of murine macrophages  
 than the wild-type strain. In addn., the bioactivity of the  
 lipopolysaccharide (LPS) of the **htrB** mutant was  
 reduced compared to that of the LPS from the parent strain as  
 measured by both a *Limulus* amoebocyte lysate endotoxin quantitation  
 assay and a tumor necrosis factor alpha bioassay. These results  
 indicated that the htrB gene plays a role in the virulence of *S.*  
*typhimurium*.

L2 ANSWER 6 OF 17 CAPLUS COPYRIGHT 1999 ACS

AN 1997:730328 CAPLUS

DN 128:21296

TI Evaluation of the virulence of nontypeable *Hemophilus influenzae* lipooligosaccharide *htrB* and *rfaD* mutants in the chinchilla model of otitis media

SO Infect. Immun. (1997), 65(11), 4431-4435

CODEN: INFIBR; ISSN: 0019-9567

AU DeMaria, T. F.; Apicella, M. A.; Nichols, W. A.; Leake, E. R.

PY 1997

AB Considerable evidence has implicated nontypeable *Hemophilus influenzae* (NTHi) lipooligosaccharide (LOS) in the pathogenesis of otitis media (OM); however, its exact role has not been conclusively established. Recently, two NTHi LOS-deficient mutants have been created and described. Strain 2019-DK1, an *rfaD* gene mutant, expresses a truncated LOS consisting of only three deoxy-D-manno-octulosonic acid residues, a single heptose, and lipid A. Strain 2019-B29, an isogenic *htrB* mutant, possesses an altered oligosaccharide core and an altered lipid A. Each strain's ability to colonize the nasopharynx and to induce OM subsequent to transbullar inoculation was evaluated in the chinchilla model. Nasopharyngeal colonization data indicate that the parent strain and both mutants are able to colonize the nasopharynx and exhibit comparable clearance kinetics. Compared with the parent and each other, however, the mutants demonstrated marked differences in virulence regarding their relative abilities to induce OM and persist in the middle ear post-transbullar inoculation. Strain B29 required a 3-log-greater dose to induce OM than the parent strain and did not exhibit evidence of sustained multiplication but persisted for the same duration as the parent. Conversely, strain-DK1, even when inoculated at a dose 4 logs greater than the parent dose, was eliminated from the middle ear 72 h after challenge. A comparison of the relative pathogenicities of these isolates provides the opportunity to address fundamental questions regarding the contribution of LOS to pathogenesis issues at the mol. level. Specifically, the impact of these LOS gene disruptions on OM pathogenesis can be defined and may thus provide potential new targets for future protection and intervention strategies.

L2 ANSWER 7 OF 17 CAPLUS COPYRIGHT 1999 ACS

AN 1997:605036 CAPLUS

DN 127:275201

TI *htrB* of *Haemophilus influenzae*: determination of biochemical activity and effects on virulence and lipooligosaccharide toxicity

SO J. Endotoxin Res. (1997), 4(3), 163-172

CODEN: JENREB; ISSN: 0968-0519

AU Nichols, W. A.; Raetz, C. R. H.; Clementz, T.; Smith, A. L.; Hanson, Searcher : Shears 308-4994

J. A.; Ketterer, M. R.; Sunshine, M.; Apicella, M. A.

PY 1997

AB The **htrB** mutant of **Haemophilus**

**influenzae** (strain B29) has been shown to lack secondary (nonhydroxylated) acyl groups in its lipid A. The authors have detd. through in vitro biochem. assays that the HtrB protein acts as a specific acyltransferase in the late stages of lipid A biosynthesis and that the preferred acyl group donor is myristoyl-acyl carrier protein. Under the conditions employed, the *Escherichia coli* precursor, Kdo2-lipid IVA, functions as a myristate acceptor. Introduction of the **Haemophilus htrB** gene into an *E. coli* mutant lacking **htrB** complements the biochem. and physiol. defects assocd. with the *E. coli htrB* mutation. Tumor necrosis factor .alpha. (TNF.alpha.) assays using murine and human macrophage cells indicated that nontypeable *H. influenzae* (NtHi) strain 2019 and *H. influenzae* type b strain A2 elicit levels of expression of TNF.alpha. that are 30-40 times greater than levels induced by the isogenic **htrB** mutants (B29 and A2B29). Studies using cell-free LOS indicated that the LOS from wild type strain 2019 elicits levels of TNF.alpha. expression that are 6-8-fold higher than those of B29. In situ hybridization studies of a primary human bronchial epithelial cell line demonstrated a greater increase of TNF.alpha. message produced in the presence of 2019 LOS than in the presence of B29 LOS. TNF.alpha. levels of the cell supernatant of cells stimulated with 2019 LOS were found to be 7-8-fold higher than levels in B29 stimulated supernatants. Using the *Limulus* amoebocyte lysate for assessment of endotoxic activity, we found that wild type LOS was 8-fold higher in endotoxic activity compared with the mutant LOS. In virulence assays using i.p. inoculation of infant rats, the **htrB** isogenic strain caused bacteremia at 50% the frequency of the wild type strain. In intranasal inoculation studies, the **htrB** mutant strain was unable to cause bacteremia whereas the wild type b parent produced bacteremia in 40-60% of the animals. These findings suggest that the **htrB** gene of *H. influenzae* is important for virulence and that host TNF.alpha. expression is attenuated in response to **htrB** mutant strains.

L2 ANSWER 8 OF 17 CAPLUS COPYRIGHT 1999 ACS

AN 1997:591347 CAPLUS

DN 127:288753

TI Mutation of the **htrB** gene in a virulent

**Salmonella typhimurium** strain by intergeneric transduction: strain construction and phenotypic characterization

SO J. Bacteriol. (1997), 179(17), 5521-5533

CODEN: JOBAAY; ISSN: 0021-9193

AU Sunshine, Melvin G.; Gibson, Bradford W.; Engstrom, Jeffrey J.; Nichols, Wade A.; Jones, Bradley D.; Apicella, Michael A.

Searcher : Shears 308-4994

PY 1997

AB The *htrB* gene product of *Haemophilus influenzae* contributes to the toxicity of the lipooligosaccharide. The *htrB* gene encodes a 2-keto-3-deoxyoctulosonic acid-dependent acyltransferase which is responsible for myristic acid substitutions at the hydroxy moiety of lipid A .beta.-hydroxymyristic acid. Mass spectroscopic anal. has demonstrated that lipid A from an *H. influenzae htrB* mutant is predominantly tetraacyl and similar in structure to lipid IVA, which has been shown to be nontoxic in animal models. We sought to construct a *Salmonella typhimurium htrB* mutant in order to investigate the contribution of *htrB* to virulence in a well-defined murine typhoid model of animal pathogenesis. To this end, an *r- m+ gale mutS recD* strain of *S. typhimurium* was constructed (MGS-7) and used in inter- and intrastrain transduction expts. with both coliphage P1 and *Salmonella* phage P22. The *Escherichia coli htrB* gene contg. a mini-Tn10 insertion was transduced from *E. coli* MLK217 into *S. typhimurium* MGS-7 via phage P1 and subsequently via phage P22 into the virulent *Salmonella* strain SL1344. All *S. typhimurium* transductants showed phenotypes similar to those described for the *E. coli htrB* mutant. Mass spectrometric anal. of the crude lipid A fraction from the lipopolysaccharide of the *S. typhimurium htrB* mutant strain showed that for the dominant hexaacyl form, a lauric acid moiety was lost at one position on the lipid A and a palmitic acid moiety was added at another position; for the less abundant heptaacyl species, the lauric acid was replaced with palmitoleic acid.

L2 ANSWER 9 OF 17 CAPLUS COPYRIGHT 1999 ACS

AN 1997:496805 CAPLUS

DN 127:107983

TI Non-toxic mutants of pathogenic gram-negative bacteria

SO PCT Int. Appl., 78 pp.

CODEN: PIXXD2

IN Apicella, Michael A.; Sunshine, Melvin G.; Lee, Na-gyong; Arumugham, Rasappa; Gibson, Bradford W.

APPLICATION NO. DATE

AI WO 96-US18984 19961127

CA 96-2238640 19961127

AU 97-11246 19961127

EP 96-942080 19961127

PATENT NO. KIND DATE APPLICATION NO. DATE

PI WO 9719688 A1 19970605 WO 96-US18984 19961127

W: AU, CA, JP, KR, MX, NZ, US, AM, AZ, BY, KG, KZ, MD, RU, TJ,

Searcher : Shears 308-4994

TM

RW: AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL,  
PT, SE

CA 2238640	AA	19970605	CA 96-2238640	19961127
AU 9711246	A1	19970619	AU 97-11246	19961127
EP 876150	A1	19981111	EP 96-942080	19961127

R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC,  
PT, IE, FI

PY 1997

1997

1997

1998

AB A method is provided for identifying, isolating, and producing **htrB mutants** of **gram-neg.** bacterial pathogens. The method comprises **mutating** the **htrB** gene of a **gram-neg.** bacterial pathogen so that there is a lack of a functional **htrB** protein, resulting in a **mutant** that lacks .gtoreq.1 secondary acyl chains contained in the wild type **gram-neg.** bacterial pathogen, and displays substantially reduced toxicity as compared to the wild type strain. The present invention also provides methods for using a vaccine formulation contg. the **htrB mutant**, the endotoxin isolated therefrom, or the endotoxin isolated therefrom which is then conjugated to a carrier protein to immunize an individual against infections caused by **gram-neg.** bacterial pathogens by administering a prophylactically effective amt. of the vaccine formulation.

L2 ANSWER 10 OF 17 CAPLUS COPYRIGHT 1999 ACS

AN 1997:279634 CAPLUS

DN 126:339596

TI Function of the Escherichia coli msbB gene, a multicopy suppressor of htrB knockouts, in the acylation of lipid A. Acylation by MsbB follows laurate incorporation by HtrB

SO J. Biol. Chem. (1997), 272(16), 10353-10360

CODEN: JBCHA3; ISSN: 0021-9258

AU Clementz, Tony; Zhou, Zhimin; Raetz, Christian R. H.

PY 1997

AB Overexpression of the Escherichia coli msbB gene on high copy plasmids suppresses the temp.-sensitive growth assocd. with **mutations** in the **htrB** gene. HtrB encodes the lauroyl transferase of lipid A biosynthesis that acylates the intermediate (Kdo)2-lipid IVA. Since msbB displays 27.5% identity and 42.2% similarity to htrB, we explored the possibility that msbB encodes a related acyltransferase. In contrast to **htrB**, exts. of strains with insertion **mutations** in msbB are not defective in transferring laurate from lauroyl acyl carrier protein to (Kdo)2-lipid IVA. However, exts. of msbB **mutants** do

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not efficiently acylate the product formed by **HtrB**, designated (Kdo)2-(lauroyl)-lipid IVA. Exts. of strains harboring **msbB+** bearing plasmids acylate (Kdo)2-(lauroyl)-lipid IVA very rapidly compared with wild type. We solubilized and partially purified **msbB** from an overproducing strain, lacking **HtrB**, **MsbB** transfers myristate or laurate, activated on ACP, to (Kdo)2-(lauroyl)-lipid IVA. Decanoyl, palmitoyl, palmitoleoyl, and (R)-3-hydroxymyristoyl-ACP are poor acyl donors. **MsbB** acylates (Kdo)2-(lauroyl)-lipid IVA .apprx.100 times faster than (Kdo)2-lipid IVA. The slow, but measurable, rate whereby **MsbB** acts on (Kdo)2-lipid IVA may explain why overexpression of **MsbB** suppresses the temp.-sensitive phenotype of **htrB** mutations. Presumably, the acyloxyacyl group generated by excess **MsbB** substitutes for the one normally formed by **HtrB**.

L2 ANSWER 11 OF 17 CAPLUS COPYRIGHT 1999 ACS

AN 1996:411262 CAPLUS

DN 125:134544

TI Mutational analysis and properties of the **msbA** gene of *Escherichia coli*, coding for an essential ABC family transporter

SO Mol. Microbiol. (1996), 20(6), 1221-1233

CODEN: MOMIEE; ISSN: 0950-382X

AU Polissi, Alessandra; Georgopoulos, Costa

PY 1996

AB The **htrB** gene was discovered because its insertional inactivation interfered with *Escherichia coli* growth and viability at temps. above 32.5.degree., as a result of accumulation of phospholipids. The **msbA** gene was originally discovered because when cloned on a low-cop-no. plasmid vector it was able to suppress the temp.-sensitive growth phenotype of an **htrB** null mutant as well as the accumulation of phospholipids. The **msbA** gene product belongs to the superfamily of ABC transporters, a universally conserved family of proteins characterized by a highly conserved ATP-binding domain. The **msbA** gene is essential for bacterial viability at all temps. To understand the physiol. role of the **MsbA** protein, we mutated the ATP-binding domain using random PCR mutagenesis. Six independent mutants were isolated and characterized. Four of these mutations resulted in single-amino-acid substitutions in non-conserved residues and were able to support cell growth at 30.degree. but not at 43.degree.. The remaining two mutations behaved as recessive lethals, and resulted in single-amino-acid substitutions in Walker motif B, one of the two highly conserved regions of the ATP-binding domain. Despite the fact that neither of these two mutant proteins can support *E. coli* growth, they both retained the ability to bind ATP in vitro. In addn., we present evidence to show that N-acetyl [3H]-glucosamine, a precursor of lipopolysaccharides, accumulates at the non-permissive temp. in the inner membrane of either **htrB** null or **msbA** conditional lethal strains. Translocation

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of the precursor to the outer membrane is restored by transformation with a plasmid contg. the wild-type *msbA* gene. A possible role for *MsbA* as a translocator of lipopolysaccharides or its precursors is discussed.

L2 ANSWER 12 OF 17 CAPLUS COPYRIGHT 1999 ACS

AN 1995:945651 CAPLUS

DN 124:25327

TI **Mutation of the *htrB* locus of**

***Haemophilus influenzae* nontypable strain 2019 is associated with modifications of lipid A and phosphorylation of the lipo-oligosaccharide**

SO J. Biol. Chem. (1995), 270(45), 27151-9

CODEN: JBCHA3; ISSN: 0021-9258

AU Lee, Na-Gyong; Sunshine, Melvin G.; Engstrom, Jeffery J.; Gibson, Bradford W.; Apicella, Michael A.

PY 1995

AB The *HtrB* protein was first identified in *Escherichia coli* as a protein required for cell viability at high temp., but its expression was not regulated by temp. An *htrB* homolog was isolated from nontypable *Haemophilus influenzae* strain (NTHi) 2019, which was able to functionally complement the *E. coli htrB* mutation. The promoter for the NTHi 2019 *htrB* gene overlaps the promoter for the *rfaE* gene, and the 2 genes are divergently transcribed. The deduced amino acid sequence of NTHi 2019 *HtrB* had 56% homol. to *E. coli HtrB*. In vitro transcription-translation anal. confirmed prodn. of a protein with an apparent mol. mass of 32-33 kDa. Primer extension anal. revealed that *htrB* was transcribed from a  $\sigma_{70}$ -dependent consensus promoter and its expression was not affected by temp. The expression of *htrB* and *rfaE* was 2.5-4-fold higher in the NTHi *htrB* mutant B29 than in the parental strain. In order to study the function of the *HtrB* protein in *Haemophilus*, 2 isogenic *htrB* mutants were generated by shuttle mutagenesis using a mini-Tn3. The *htrB* mutants initially showed temp. sensitivity, but they lost the sensitivity after a few passages at 30.degree. and were able to grow at 37.degree.. They also showed hypersensitivity to deoxycholate and kanamycin, which persisted on passage. SDS-PAGE anal. revealed that the lipo-oligosaccharide (LOS) isolated from these mutants migrated faster than the wild type LOS and its color changed from black to brown as has been described for *E. coli htrB* mutants. Immunoblotting anal. also showed that the LOS from the *htrB* mutants lost reactivity to a monoclonal antibody, 6E4, which binds to the wild type NTHi 2019 LOS. Electrospray ionization-mass spectrometry anal. of the O-deacylated LOS oligosaccharide indicated a modification of the core structure characterized in part by a net loss in

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phosphoethanolamine. Mass spectrometric anal. of the lipid A of the **htrB mutant** indicated a loss of one or both myristic acid substitutions. These data suggest that HtrB is a multifunctional protein and may play a controlling role in regulating cell responses to various environmental changes.

L2 ANSWER 13 OF 17 CAPLUS COPYRIGHT 1999 ACS

AN 1993:487844 CAPLUS

DN 119:87844

TI The essential *Escherichia coli* **msbA** gene, a multicopy suppressor of null mutations in the **htrB** gene, is related to the universally conserved family of ATP-dependent translocators

SO Mol. Microbiol. (1993), 7(1), 69-79

CODEN: MOMIEE; ISSN: 0950-382X

AU Karow, Margaret; Georgopoulos, Costa

PY 1993

AB The **msbA** gene, isolated as a multicopy suppressor of the HtrB temp.-sensitive phenotype, was characterized. The **msbA** gene maps to 20.5 min on the *Escherichia coli* genetic map and encodes a protein with an estd. mol. mass of 64,460 Da, with the properties of an integral membrane protein. The amino acid sequence of **MsbA** is very similar to those of the family of ATP-dependent translocators, which includes the hemolysin B protein of *E. coli* and the mammalian multidrug resistance (MDR) proteins. Mutational anal. of **msbA** indicates that it may form an operon with a down-stream gene, **orfe**, and that both of these genes are essential for bacterial viability under all growth conditions tested.

L2 ANSWER 14 OF 17 CAPLUS COPYRIGHT 1999 ACS

AN 1993:162023 CAPLUS

DN 118:162023

TI Isolation and characterization of the *Escherichia coli* **msbB** gene, a multicopy suppressor of null mutations in the high-temperature requirement gene **htrB**

SO J. Bacteriol. (1992), 174(3), 702-10

CODEN: JOBAAY; ISSN: 0021-9193

AU Karow, Margaret; Georgopoulos, Costa

PY 1992

AB Previous work established that the **htrB** gene of *E. coli* is required for growth in rich media at temps. about 32.5.degree. but not at lower temps. In an effort to det. the functional role of the **htrB** gene product, the authors isolated a multicopy suppressor of **htrB**, called **msbB**. The **msbB** gene has been mapped to 40.5 min on the *E. coli* genetic map, in a 12- to 15-kb gap of the genomic library made by Y. Kohara et al. (1987). Mapping data show that the order of genes in the region is **eda-edd-zwf-pykA-msbB**. The **msbB** gene codes for a protein of 37,410 Da whose amino acid sequence is similar to that of HtrB and, like HtrB, the protein is very basic in

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nature. The similarity of the HtrB and MsbB proteins may indicate that they play functionally similar roles. **Mutational** anal. of msbB shows that the gene is not essential for *E. coli* growth; however, the *htrB* msbB double **mutant** exhibits a unique morphol. phenotype at 30.degree. not seen with either of the single **mutants**. Anal. of both msbB and *htrB* **mutants** shows that these bacteria are resistant to 4-fold more deoxycholate than are wild-type bacteria but not to other hydrophobic substances. The addn. of quaternary ammonium compds. rescues the temp.-sensitive phenotype of *htrB* bacteria, and this rescue is abolished by the simultaneous addn. of Mg<sup>2+</sup> or Ca<sup>2+</sup>. These results suggest that MsbB and HtrB play an important role in outer membrane structure and/or function.

L2 ANSWER 15 OF 17 CAPLUS COPYRIGHT 1999 ACS

AN 1993:19016 CAPLUS

DN 118:19016

TI The lethal phenotype caused by null **mutations** in the *Escherichia coli htrB* gene is suppressed by **mutations** in the accBC operon, encoding two subunits of acetyl coenzyme A carboxylase

SO J. Bacteriol. (1992), 174(22), 7407-18

CODEN: JOBAAY; ISSN: 0021-9193

AU Karow, Margaret; Fayet, Olivier; Georgopoulos, Costa

PY 1992

AB Insertion **mutations** in the *E. coli htrB*

gene result in the unique phenotype of not affecting growth at temps. <32.5.degree. but leading to a loss of viability at temps. above this in rich media. When *htrB* bacteria growing in rich media were shifted to the nonpermissive temp. of 42.degree., they continued to grow at a rate similar to that of 30.degree. but they produced phospholipids at the rate required for growth at 42.degree.. This led to the accumulation of more than twice as much phospholipid per mg of protein compared with that in wild-type bacteria. Consistent with *HtrB* playing a role in phospholipid biosynthesis, one complementation group of spontaneously arising **mutations** that suppressed *htrB*-induced lethality were mapped to the accBC operon. This operon codes for the biotin carboxyl carrier protein and biotin carboxylase subunits of the acetyl CoA carboxylase enzyme complex, which catalyzes the 1st step in fatty acid biosynthesis. Four suppressor mutations mapped to this operon. Two alleles were identified as mutations in the accC gene, the 3rd allele was identified as a mutation in the accB gene, and the 4th as an insertion of an IS1 transposable element in the promoter region of the operon, resulting in reduced transcription. The suppressor **mutations** caused a decrease in the rate of phospholipid biosynthesis, restoring the balance between the biosynthesis of phospholipids and growth rate, thus enabling *htrB* bacteria

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to grow at high temps.

L2 ANSWER 16 OF 17 CAPLUS COPYRIGHT 1999 ACS

AN 1992:525535 CAPLUS

DN 117:125535

TI Sequencing, mutational analysis, and transcriptional regulation of the *Escherichia coli* htrB gene

SO Mol. Microbiol. (1991), 5(9), 2285-92

CODEN: MOMIEE; ISSN: 0950-382X

AU Karow, M.; Georgopoulos, C.

PY 1991

AB The *E. coli* htrB gene was originally discovered because its insertional inactivation led to an exquisitely temp.-sensitive phenotype in rich media, i.e. the ability to form colonies at temps. below 32.degree., but not above 33.degree.. The htrB gene has been sequenced. It can potentially code for 2 proteins, with Mr values of 35407 Da and 8669 Da, that are encoded by overlapping, divergent open reading frames. The data are consistent with the 35407 Da protein being HtrB. Northern blot anal. clearly shows that the monocistronic htrB message is not under heat-shock regulation. The flanking DNA was also sequenced and a new gene, designated orf39.9, located immediately adjacent to htrB, but divergently transcribed was discovered.

L2 ANSWER 17 OF 17 CAPLUS COPYRIGHT 1999 ACS

AN 1992:102492 CAPLUS

DN 116:102492

TI Complex phenotypes of null mutations in the htr genes, whose products are essential for *Escherichia coli* growth at elevated temperatures

SO Res. Microbiol. (1991), 142(2-3), 289-94

CODEN: RMCREW; ISSN: 0923-2508

AU Karow, M.; Raina, S.; Georgopoulos, C.; Fayet, O.

PY 1991

AB Transposon insertion, followed by screening, has allowed the identification of a set of genes, called htr, whose products are required for *E. coli* growth at elevated temps. The htrB gene maps at 23.5 min on the *E. coli* genetic map. It codes for a very basic, hydrophobic, 35,000-Mr polypeptide, possessing a putative membrane-spanning domain. At nonpermissive temp., htrB mutant bacteria stop dividing, followed by the formation of bulges and eventual lysis. The htrC gene maps at 90 min, is under  $\sigma_{32}$  regulation, and codes for a 21,130-Mr polypeptide. At 43.degree. htrC mutant bacteria gradually lyse, whereas at intermediate temps. they filament extensively. Finally, the htrM gene maps at 81 min, is under  $\sigma_{32}$  regulation, and codes for a 35,000-Mr polypeptide. The HtrM null phenotype included inability to grow at >42.degree., extreme mucoidness, and sensitivity to bile salts, even at the permissive

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temps. The htrM gene is identical to the rfaD gene, whose product is required for the biosynthesis of the lipopolysaccharide precursor ADP-L-glycero-D-mannoheptose.

=> d his 13- ful; d 1-21 bib abs

(FILE 'MEDLINE, BIOSIS, EMBASE, LIFESCI, BIOTECHDS, WPIDS, CONFSCI, SCISEARCH, JICST-EPLUS, PROMT, TOXLIT, TOXLINE, DRUGU, DRUGNL, DRUGB' ENTERED AT 10:26:29 ON 01 MAR 1999)

L3 98 SEA ABB=ON PLU=ON L1  
 L4 98 SEA ABB=ON PLU=ON L3 AND (GRAM(W) (NEGATIVE OR NEG) OR SALMONELL? OR COLI OR HAEMOPHIL? OR HEMOPHIL?)  
 L5 21 DUP REM L4 (77 DUPLICATES REMOVED)

L5 ANSWER 1 OF 21 BIOTECHDS COPYRIGHT 1999 DERWENT INFORMATION LTD  
 AN 98-10001 BIOTECHDS  
 TI New non-functional mutant nucleic acid from *Salmonella* msbB or htrB genes;  
 lipid-A preparation by *Salmonella* typhimurium msbB, htrB mutant gene transfer and expression in host cell, used for e.g. septic shock therapy, gene therapy, or recombinant vaccine

AU Maskell D J; Dougan G  
 PA Univ.London  
 LO London, UK.  
 PI WO 9833923 6 Aug 1998  
 AI WO 98-GB291 30 Jan 1998  
 PRAI GB 97-1887 30 Jan 1997; GB 97-1886 30 Jan 1997  
 DT Patent  
 LA English  
 OS WPI: 98-437476 [37]  
 AN 98-10001 BIOTECHDS  
 AB A new *Salmonella* bp. msbB or htrB gene sequence has a mutation that results in the loss of the encoded protein or its activity, and results in a lipid-A molecule of reduced toxicity. Also new are protein encoded by the DNA, a recombinant DNA construct containing the DNA, and a microorganism containing the mutated msbB and/or htrB gene, where one or both of the genes are inactivated. The microorganisms may be used as live vaccines to protect against infection by the corresponding wild-type, in humans, mammals or birds. The microorganisms may also be used for production of nucleic acid or proteins for therapy or gene therapy, e.g. lipopolysaccharides may be used as endotoxin antagonists for treatment of septic shock. The new DNA is preferably a mutant msbB gene derived from *Salmonella* typhimurium C5 (NCIMB 40856), but may also be from one of 18 specified *Salmonella* spp. or from one of 19 specified genera. The mutant is produced by inserting a kanamycin-resistance cassette into the gene, conjugating as a

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suicide vector into a recipient that is to be **mutated**,  
and transducing into another recipient. (38pp)

L5 ANSWER 2 OF 21 MEDLINE DUPLICATE 2  
AN 1998241619 MEDLINE  
DN 98241619  
TI Function of *Escherichia coli* MsbA, an essential ABC family  
transporter, in lipid A and phospholipid biosynthesis.  
AU Zhou Z; White K A; Polissi A; Georgopoulos C; Raetz C R  
CS Department of Biochemistry, Duke University Medical Center, Durham,  
North Carolina 27710, USA.  
NC GM-51310 (NIGMS)  
SO JOURNAL OF BIOLOGICAL CHEMISTRY, (1998 May 15) 273 (20) 12466-75.  
Journal code: HIV. ISSN: 0021-9258.  
CY United States  
DT Journal; Article; (JOURNAL ARTICLE)  
LA English  
FS Priority Journals; Cancer Journals  
EM 199808  
EW 19980803  
AB The *Escherichia coli* msbA gene, first identified as a  
multicopy suppressor of **htrB mutations**, has been  
proposed to transport nascent core-lipid A molecules across the  
inner membrane (Polissi, A., and Georgopoulos, C. (1996) Mol.  
Microbiol. 20, 1221-1233). msbA is an essential *E. coli*  
gene with high sequence similarity to mammalian Mdr proteins and  
certain types of bacterial ABC transporters. **htrB** is  
required for growth above 32 degreesC and encodes the  
lauroyltransferase that acts after Kdo addition during lipid A  
biosynthesis (Clementz, T., Bednarski, J., and Raetz, C. R. H.  
(1996) J. Biol. Chem. 271, 12095-12102). By using a quantitative new  
32Pi labeling technique, we demonstrate that hexa-acylated species  
of lipid A predominate in the outer membranes of wild type *E.*  
*coli* labeled for several generations at 42 degreesC. In  
contrast, in **htrB mutants** shifted to 42 degreesC  
for 3 h, tetra-acylated lipid A species and glycerophospholipids  
accumulate in the inner membrane. Extra copies of the cloned msbA  
gene restore the ability of **htrB mutants** to grow  
at 42 degreesC, but they do not increase the extent of lipid A  
acylation. However, a significant fraction of the tetra-acylated  
lipid A species that accumulate in **htrB mutants**  
are transported to the outer membrane in the presence of extra  
copies of msbA. *E. coli* strains in which msbA synthesis is  
selectively shut off at 42 degreesC accumulate hexa-acylated lipid A  
and glycerophospholipids in their inner membranes. Our results  
support the view that MsbA plays a role in lipid A and possibly  
glycerophospholipid transport. The tetra-acylated lipid A precursors  
that accumulate in **htrB mutants** may not be  
transported as efficiently by MsbA as are penta- or hexa-acylated

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lipid A species.

L5 ANSWER 3 OF 21 MEDLINE DUPLICATE 3  
 AN 1998340543 MEDLINE  
 DN 98340543  
 TI Role of the O-antigen of lipopolysaccharide, and possible roles of growth rate and of NADH:ubiquinone oxidoreductase (nuo) in competitive tomato root-tip colonization by *Pseudomonas fluorescens* WCS365.  
 AU Dekkers L C; van der Bij A J; Mulders I H; Phoelich C C; Wentwoord R A; Glandorf D C; Wijffelman C A; Lugtenberg B J  
 CS Leiden University, Institute of Molecular Plant Sciences, Clusius Laboratory, The Netherlands.. Dekkers@rulbim.leidenuniv.nl  
 SO MOLECULAR PLANT-MICROBE INTERACTIONS, (1998 Aug) 11 (8) 763-71. Journal code: A9P. ISSN: 0894-0282.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals  
 OS GENBANK-Y14568; GENBANK-Y14569  
 EM 199810  
 EW 19981004  
 AB Colonization-defective, transposon-induced **mutants** of the efficient root colonizer *Pseudomonas fluorescens* WCS365 were identified with a gnotobiotic system. Most **mutants** were impaired in known colonization traits, i.e., prototrophy for amino acids, motility, and synthesis of the O-antigen of LPS (lipopolysaccharide). **Mutants** lacking the O-antigen of LPS were impaired in both colonization and competitive growth whereas one **mutant** (PCL1205) with a shorter O-antigen chain was defective only in colonization ability, suggesting a role for the intact O-antigen of LPS in colonization. Eight competitive colonization **mutants** that were not defective in the above-mentioned traits colonized the tomato root tip well when inoculated alone, but were defective in competitive root colonization of tomato, radish, and wheat, indicating they contained **mutations** affecting host range. One of these eight **mutants** (PCL1201) was further characterized and contains a **mutation** in a gene that shows homology to the *Escherichia coli* nuo4 gene, which encodes a subunit of one of two known NADH:ubiquinone oxidoreductases. Competition experiments in an oxygen-poor medium between **mutant** PCL1201 and its parental strain showed a decreased growth rate of **mutant** PCL1201. The requirement of the nuo4 gene homolog for optimal growth under conditions of oxygen limitation suggests that the root-tip environment is micro-aerobic. A **mutant** characterized by a slow growth rate (PCL1216) was analyzed further and contained a **mutation** in a gene with similarity to the *E. coli* HtrB protein, a lauroyl transferase that functions in lipid  
 Searcher : Shears 308-4994

## A biosynthesis.

L5 ANSWER 4 OF 21 BIOTECHDS COPYRIGHT 1999 DERWENT INFORMATION LTD  
 AN 97-08966 BIOTECHDS  
 TI New **Gram-negative** bacterial pathogen vaccines;  
     **Gram-negative** bacterium **htrB**  
     endotoxin gene **mutagenesis** for reduced toxicity and  
     use as a vaccine  
 AU Apicella M A; Sunshine M G; Lee N G; Arumugham R; Gibson B W  
 PA Univ.Iowa-Res.Found.; Univ.California; American-Cyanamid  
 LO Iowa City, IA, USA; Oakland, CA, USA; Madison, NJ, USA.  
 PI WO 9719688 5 Jun 1997  
 AI WO 96-US18984 27 Nov 1996  
 PRAI US 95-565943 1 Dec 1995  
 DT Patent  
 LA English  
 OS WPI: 97-310355 [28]  
 AN 97-08966 BIOTECHDS  
 AB A method for immunizing an individual to prevent disease caused by  
     a **Gram-negative** bacterial pathogen is claimed,  
     which involves vaccinating the individual with a formulation  
     (claimed) consisting of a **Gram-negative**  
     bacterium **htrB mutant**, endotoxin isolated from  
     the **mutant**, endotoxin isolated from the **mutant**  
     and conjugated with a carrier protein, or a **mutant** which  
     has been genetically engineered to express at least one  
     heterologous vaccine antigen as the active ingredient. Also  
     claimed are methods for producing a **mutant** endotoxin or a  
     **Gram-negative** bacterium **mutant** having  
     substantially reduced toxicity as compared with the wild-type  
     endotoxin or bacterium, which involves **mutating** an  
     **htrB** gene within the bacterium causing a phenotype  
     characterized by a **mutant** endotoxin lacking at least one  
     secondary acyl chain on lipid-A contained in the wild-type  
     bacterium. The endotoxins have reduced toxicity compared with the  
     wild-type endotoxins and yet retain antigenicity. The compositions  
     can be used as prophylactic or therapeutic vaccines against  
     endotoxic shock and **Gram-negative** bacteremia.  
     (79pp)

L5 ANSWER 5 OF 21 MEDLINE  
 AN 97256743 MEDLINE  
 DN 97256743  
 TI Function of the Escherichia coli msbB gene, a multicopy  
     suppressor of htrB knockouts, in the acylation of lipid A. Acylation  
     by MsbB follows laurate incorporation by HtrB.  
 AU Clementz T; Zhou Z; Raetz C R  
 CS Department of Biochemistry, Duke University Medical Center, Durham,  
     North Carolina 27710, USA.

Searcher : Shears 308-4994

NC GM-51310 (NIGMS)  
 SO JOURNAL OF BIOLOGICAL CHEMISTRY, (1997 Apr 18) 272 (16) 10353-60.  
 Journal code: HIV. ISSN: 0021-9258.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals; Cancer Journals  
 EM 199707  
 EW 19970703  
 AB Overexpression of the *Escherichia coli* msbB gene on high copy plasmids suppresses the temperature-sensitive growth associated with mutations in the htrB gene. htrB encodes the lauroyl transferase of lipid A biosynthesis that acylates the intermediate (Kdo)2-lipid IVA (Brozek, K. A., and Raetz, C. R. H. (1990) J. Biol. Chem. 265, 15410-15417). Since msbB displays 27.5% identity and 42.2% similarity to htrB, we explored the possibility that msbB encodes a related acyltransferase. In contrast to htrB, extracts of strains with insertion mutations in msbB are not defective in transferring laurate from lauroyl acyl carrier protein to (Kdo)2-lipid IVA. However, extracts of msbB mutants do not efficiently acylate the product formed by HtrB, designated (Kdo)2-(lauroyl)-lipid IVA. Extracts of strains harboring msbB+ bearing plasmids acylate (Kdo)2-(lauroyl)-lipid IVA very rapidly compared with wild type. We solubilized and partially purified MsbB from an overproducing strain, lacking HtrB. MsbB transfers myristate or laurate, activated on ACP, to (Kdo)2-(lauroyl)-lipid IVA. Decanoyl, palmitoyl, palmitoleoyl, and (R)-3-hydroxymyristoyl-ACP are poor acyl donors. MsbB acylates (Kdo)2-(lauroyl)-lipid IVA about 100 times faster than (Kdo)2-lipid IVA. The slow, but measurable, rate whereby MsbB acts on (Kdo)2-lipid IVA may explain why overexpression of MsbB suppresses the temperature-sensitive phenotype of htrB mutations. Presumably, the acyloxyacyl group generated by excess MsbB substitutes for the one normally formed by HtrB.

L5 ANSWER 6 OF 21 MEDLINE DUPLICATE 6  
 AN 97431504 MEDLINE  
 DN 97431504  
 TI Mutation of the htrB gene in a virulent *Salmonella typhimurium* strain by intergeneric transduction: strain construction and phenotypic characterization.  
 AU Sunshine M G; Gibson B W; Engstrom J J; Nichols W A; Jones B D; Apicella M A  
 CS Department of Microbiology, The University of Iowa, Iowa City 52242, USA.  
 NC AI38268 (NIAID)  
 AI24616 (NIAID)  
 AI31254 (NIAID)



+  
 SO JOURNAL OF BACTERIOLOGY, (1997 Sep) 179 (17) 5521-33.  
 Journal code: HH3. ISSN: 0021-9193.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals  
 EM 199712  
 AB The **htrB** gene product of *Haemophilus influenzae* contributes to the toxicity of the lipooligosaccharide. The **htrB** gene encodes a 2-keto-3-deoxyoctulosonic acid-dependent acyltransferase which is responsible for myristic acid substitutions at the hydroxy moiety of lipid A beta-hydroxymyristic acid. Mass spectroscopic analysis has demonstrated that lipid A from an *H. influenzae htrB* mutant is predominantly tetraacyl and similar in structure to lipid IV(A), which has been shown to be nontoxic in animal models. We sought to construct a *Salmonella typhimurium htrB* mutant in order to investigate the contribution of **htrB** to virulence in a well-defined murine typhoid model of animal pathogenesis. To this end, an r- m+ galE mutS recD strain of *S. typhimurium* was constructed (MGS-7) and used in inter- and intrastrain transduction experiments with both coliphage P1 and *Salmonella* phage P22. The *Escherichia coli htrB* gene containing a mini-Tn10 insertion was transduced from *E. coli* MLK217 into *S. typhimurium* MGS-7 via phage P1 and subsequently via phage P22 into the virulent *Salmonella* strain SL1344. All *S. typhimurium* transductants showed phenotypes similar to those described for the *E. coli htrB* mutant. Mass spectrometric analysis of the crude lipid A fraction from the lipopolysaccharide of the *S. typhimurium htrB* mutant strain showed that for the dominant hexaacyl form, a lauric acid moiety was lost at one position on the lipid A and a palmitic acid moiety was added at another position; for the less abundant heptaacyl species, the lauric acid was replaced with palmitoleic acid.

L5 ANSWER 7 OF 21 MEDLINE DUPLICATE 7  
 AN 1998013113 MEDLINE  
 DN 98013113  
 TI Study of the role of the **htrB** gene in *Salmonella typhimurium* virulence.  
 AU Jones B D; Nichols W A; Gibson B W; Sunshine M G; Apicella M A  
 CS Department of Microbiology, University of Iowa College of Medicine, Iowa City 52242-1109, USA.. bjones@blue.weeg.uiowa.edu  
 NC AI38268 (NIAID)  
 RR01614 (NCRR)  
 AI 31254 (NIAID)

+

Searcher : Shears 308-4994

SO INFECTION AND IMMUNITY, (1997 Nov) 65 (11) 4778-83.  
Journal code: GO7. ISSN: 0019-9567.

CY United States

DT Journal; Article; (JOURNAL ARTICLE)

LA English

FS Priority Journals; Cancer Journals

EM 199801

EW 19980104

AB We have undertaken a study to investigate the contribution of the **htrB** gene to the virulence of pathogenic **Salmonella typhimurium**. An **htrB::mini-Tn10** mutation from **Escherichia coli** was transferred by transduction to the mouse-virulent strain **S. typhimurium** SL1344 to create an **htrB** mutant. The **S. typhimurium htrB** mutant was inoculated into mice and found to be severely limited in its ability to colonize organs of the lymphatic system and to cause systemic disease in mice. A variety of experiments were performed to determine the possible reasons for this loss of virulence. Serum killing assays revealed that the **S. typhimurium htrB** mutant was as resistant to killing by complement as the wild-type strain. However, macrophage survival assays revealed that the **S. typhimurium htrB** mutant was more sensitive to the intracellular environment of murine macrophages than the wild-type strain. In addition, the bioactivity of the lipopolysaccharide (LPS) of the **htrB** mutant was reduced compared to that of the LPS from the parent strain as measured by both a *Limulus* amoebocyte lysate endotoxin quantitation assay and a tumor necrosis factor alpha bioassay. These results indicate that the **htrB** gene plays a role in the virulence of **S. typhimurium**.

L5 ANSWER 8 OF 21 MEDLINE

DUPLICATE 8

AN 1998013065 MEDLINE

DN 98013065

TI Evaluation of the virulence of nontypeable **Haemophilus influenzae** lipooligosaccharide **htrB** and **rfaD** mutants in the chinchilla model of otitis media.

AU DeMaria T F; Apicella M A; Nichols W A; Leake E R

CS The Ohio State University, Columbus 43210, USA..  
tdemaria@pop.service.acs.ohiostate.edu

NC 5 R01 DC00090-24 (NIDCD)  
R01 A124616-08

SO INFECTION AND IMMUNITY, (1997 Nov) 65 (11) 4431-5.  
Journal code: GO7. ISSN: 0019-9567.

CY United States

DT Journal; Article; (JOURNAL ARTICLE)

LA English

FS Priority Journals; Cancer Journals

EM 199801

Searcher : Shears 308-4994

EW 19980104

AB Considerable evidence has implicated nontypeable *Haemophilus influenzae* (NTHi) lipooligosaccharide (LOS) in the pathogenesis of otitis media (OM); however, its exact role has not been conclusively established. Recently, two NTHi LOS-deficient **mutants** have been created and described. Strain 2019-DK1, an *rfaD* gene **mutant**, expresses a truncated LOS consisting of only three deoxy-D-manno-octulosonic acid residues, a single heptose, and lipid A. Strain 2019-B29, an isogenic *htrB* **mutant**, possesses an altered oligosaccharide core and an altered lipid A. Each strain's ability to colonize the nasopharynx and to induce OM subsequent to transbullar inoculation was evaluated in the chinchilla model. Nasopharyngeal colonization data indicate that the parent strain and both **mutants** are able to colonize the nasopharynx and exhibit comparable clearance kinetics. Compared with the parent and each other, however, the **mutants** demonstrated marked differences in virulence regarding their relative abilities to induce OM and persist in the middle ear post-transbullar inoculation. Strain B29 required a 3-log-greater dose to induce OM than the parent strain and did not exhibit evidence of sustained multiplication but persisted for the same duration as the parent. Conversely, strain-DK1, even when inoculated at a dose 4 logs greater than the parent dose, was eliminated from the middle ear 72 h after challenge. A comparison of the relative pathogenicities of these isolates provides the opportunity to address fundamental questions regarding the contribution of LOS to pathogenesis issues at the molecular level. Specifically, the impact of these LOS gene disruptions on OM pathogenesis can be defined and may thus provide potential new targets for future protection and intervention strategies.

L5 ANSWER 9 OF 21 BIOSIS COPYRIGHT 1999 BIOSIS DUPLICATE 9

AN 1997:440316 BIOSIS

DN PREV199799739519

TI *HtrB* of *Haemophilus influenzae*: Determination of biochemical activity and effects on virulence and lipooligosaccharide toxicity.

AU Nichols, W. A.; Raetz, C. R. H.; Clementz, T.; Smith, A. L.; Hanson, J. A.; Ketterer, M. R.; Sunshine, M.; Apicella, M. A. (1)

CS (1) Dep. Microbiol. BSB-3-403, Univ. Iowa Coll. Med., 51 Newton Road, Iowa City, IA 52242 USA

SO Journal of Endotoxin Research, (1997) Vol. 4, No. 3, pp. 163-172. ISSN: 0968-0519.

DT Article

LA English

AB The *htrB* **mutant** of *Haemophilus*

*influenzae* (strain B29) has been shown to lack secondary (nonhydroxylated) acyl groups in its lipid A. We have determined through in vitro biochemical assays that the *HtrB* protein

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acts as a specific acyltransferase in the late stages of lipid A biosynthesis and that the preferred acyl group donor is myristoyl-acyl carrier protein. Under the conditions employed, the *Escherichia coli* precursor, Kdo-2-lipid IV-A, functions as a myristate acceptor. Introduction of the *Haemophilus htrB* gene into an *E. coli* mutant lacking *htrB* complements the biochemical and physiological defects associated with the *E. coli htrB* mutation. Tumor necrosis factor alpha (TNF-alpha) assays using murine and human macrophage cells indicated that nontypeable *H. influenzae* (NtHi) strain 2019 and *H. influenzae* type b strain A2 elicit levels of expression of TNF-alpha that are 30-40 times greater than levels induced by the isogenic *htrB* mutants (B29 and A2B29). Studies using cell-free LOS indicated that the LOS from wild type strain 2019 elicits levels of TNF-alpha expression that are 6-8-fold higher than those of B29. In situ hybridization studies of a primary human bronchial epithelial cell line demonstrated a greater increase of TNF-alpha message produced in the presence of 2019 LOS than in the presence of B29 LOS. TNF-alpha levels of the cell supernatant of cells stimulated with 2019 LOS were found to be 7-8-fold higher than levels in B29 stimulated supernatants. Using the *Limulus amoebocyte* lysate for assessment of endotoxic activity, we found that wild type LOS was 8-fold higher in endotoxic activity compared with the mutant LOS. In virulence assays using intraperitoneal inoculation of infant rats, the *htrB* isogenic strain caused bacteremia at 50% the frequency of the wild type strain. In intranasal inoculation studies, the *htrB* mutant strain was unable to cause bacteremia whereas the wild type b parent produced bacteremia in 40-60% of the animals. These findings suggest that the *htrB* gene of *H. influenzae* is important for virulence and that host TNF-alpha expression is attenuated in response to *htrB* mutant strains.

L5 ANSWER 10 OF 21 MEDLINE DUPLICATE 10  
 AN 96405645 MEDLINE  
 DN 96405645  
 TI Mutational analysis and properties of the *msbA* gene of *Escherichia coli*, coding for an essential ABC family transporter.  
 AU Polissi A; Georgopoulos C  
 CS Department de Biochimie Medicale, Centre Medical Universitaire, Geneva, Switzerland.. AP44783@ggr.co.uk  
 SO MOLECULAR MICROBIOLOGY, (1996 Jun) 20 (6) 1221-33.  
 Journal code: MOM. ISSN: 0950-382X.  
 CY ENGLAND: United Kingdom  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals  
 EM 199704

Searcher : Shears 308-4994

EW 19970404

AB The **htrB** gene was discovered because its insertional inactivation interfered with *Escherichia coli* growth and viability at temperatures above 32.5 degrees C, as a result of accumulation of phospholipids. The **msbA** gene was originally discovered because when cloned on a low-copy-number plasmid vector it was able to suppress the temperature-sensitive growth phenotype of an **htrB** null mutant as well as the accumulation of phospholipids. The **msbA** gene product belongs to the superfamily of ABC transporters, a universally conserved family of proteins characterized by a highly conserved ATP-binding domain. The **msbA** gene is essential for bacterial viability at all temperatures. In order to understand the physiological role of the **MsbA** protein, we mutated the ATP-binding domain using random PCR mutagenesis. Six independent mutants were isolated and characterized. Four of these mutations resulted in single-amino-acid substitutions in non-conserved residues and were able to support cell growth at 30 degrees C but not at 43 degrees C. The remaining two mutations behaved as recessive lethals, and resulted in single-amino-acid substitutions in Walker motif B, one of the two highly conserved regions of the ATP-binding domain. Despite the fact that neither of these two mutant proteins can support *E. coli* growth, they both retained the ability to bind ATP in vitro. In addition, we present evidence to show that N-acetyl [3H]-glucosamine, a precursor of lipopolysaccharides, accumulates at the non-permissive temperature in the inner membrane of either **htrB** null or **msbA** conditional lethal strains. Translocation of the precursor to the outer membrane is restored by transformation with a plasmid containing the wild-type **msbA** gene. A possible role for **MsbA** as a translocator of lipopolysaccharides or its precursors is discussed.

L5 ANSWER 11 OF 21 BIOSIS COPYRIGHT 1999 BIOSIS

AN 1996:259716 BIOSIS

DN PREV199698815845

TI **Haemophilus influenzae htrB mutants**

induce a reduced production of tumor necrosis factor by mouse macrophage-like cells.

AU Nichols, Wade A. (1); Sunshine, Melvin G. (1); Harty, John T. (1); Smith, Arnold L.; Apicella, Michael A. (1)

CS (1) Univ. Iowa, Iowa City, IA USA

SO Abstracts of the General Meeting of the American Society for Microbiology, (1996) Vol. 96, No. 0, pp. 232.

Meeting Info.: 96th General Meeting of the American Society for Microbiology New Orleans, Louisiana, USA May 19-23, 1996

ISSN: 1060-2011.

DT Conference

LA English

L5 ANSWER 12 OF 21 MEDLINE  
 AN 96070820 MEDLINE  
 DN 96070820  
 TI **Mutation of the htrB locus of**  
**Haemophilus influenzae nontypable strain 2019 is associated**  
**with modifications of lipid A and phosphorylation of the**  
**lipo-oligosaccharide.**  
 AU Lee N G; Sunshine M G; Engstrom J J; Gibson B W; Apicella M A  
 CS Department of Microbiology, University of Iowa, Iowa City 52242,  
 USA..  
 NC AI 24616 (NIAID)  
 NCRR B RTP 01614  
 SO JOURNAL OF BIOLOGICAL CHEMISTRY, (1995 Nov 10) 270 (45) 27151-9.  
 Journal code: HIV. ISSN: 0021-9258.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals; Cancer Journals  
 OS GENBANK-U17642  
 EM 199602  
 AB The **HtrB** protein was first identified in *Escherichia coli* as a protein required for cell viability at high temperature, but its expression was not regulated by temperature. We isolated an **htrB** homologue from non-typable *Haemophilus influenzae* strain (NTHi) 2019, which was able to functionally complement the *E. coli htrB* mutation. The promoter for the NTHi 2019 **htrB** gene overlaps the promoter for the *rfaE* gene, and the two genes are divergently transcribed. The deduced amino acid sequence of NTHi 2019 **HtrB** had 56% homology to *E. coli* **HtrB**. In vitro transcription-translation analysis confirmed production of a protein with an apparent molecular mass of 32-33 kDa. Primer extension analysis revealed that **htrB** was transcribed from a sigma 70-dependent consensus promoter and its expression was not affected by temperature. The expression of **htrB** and *rfaE* was 2.5-4 times higher in the NTHi **htrB** mutant B29 than in the parental strain. In order to study the function of the **HtrB** protein in *Haemophilus*, we generated two isogenic **htrB** mutants by shuttle mutagenesis using a mini-Tn3. The **htrB** mutants initially showed temperature sensitivity, but they lost the sensitivity after a few passages at 30 degrees C and were able to grow at 37 degrees C. They also showed hypersensitivity to deoxycholate and kanamycin, which persisted on passage. SDS-polyacrylamide gel electrophoresis analysis revealed that the lipo-oligosaccharide (LOS) isolated from these mutants migrated faster than the wild type LOS and its color changed from black to brown as has been described for *E. coli htrB* mutants. Immunoblotting

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analysis also showed that the LOS from the **htrB** mutants lost reactivity to a monoclonal antibody, 6E4, which binds to the wild type NTHi 2019 LOS. Electrospray ionization-mass spectrometry analysis of the O-deacylated LOS oligosaccharide indicated a modification of the core structure characterized in part by a net loss in phosphoethanolamine. Mass spectrometric analysis of the lipid A of the **htrB** mutant indicated a loss of one or both myristic acid substitutions. These data suggest that **HtrB** is a multifunctional protein and may play a controlling role in regulating cell responses to various environmental changes.

L5 ANSWER 13 OF 21 MEDLINE  
 AN 95172727 MEDLINE  
 DN 95172727  
 TI Molecular cloning and characterization of the nontypeable **Haemophilus influenzae** 2019 rfaE gene required for lipopolysaccharide biosynthesis.  
 AU Lee N G; Sunshine M G; Apicella M A  
 CS Department of Microbiology, University of Iowa, Iowa City 52242..  
 NC AI 24616 (NIAID)  
 SO INFECTION AND IMMUNITY, (1995 Mar) 63 (3) 818-24.  
 Journal code: GO7. ISSN: 0019-9567.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals; Cancer Journals  
 OS GENBANK-U17642  
 EM 199506  
 AB The lipooligosaccharide (LOS) of nontypeable **Haemophilus influenzae** (NTHi) is an important factor in pathogenesis and virulence. In an attempt to elucidate the genes involved in LOS biosynthesis, we have cloned the rfaE gene from NTHi 2019 by complementing a **Salmonella typhimurium** rfaE mutant strain with an NTHi 2019 plasmid library. The rfaE mutant synthesizes lipopolysaccharide (LPS) lacking heptose, and the rfaE gene is postulated to be involved in ADP-heptose synthesis. Retransformation with the plasmid containing 4 kb of NTHi DNA isolated from a reconstituted mutant into rfaE mutants gave wild-type LPS phenotypes. Sodium dodecyl sulfate-polyacrylamide gel electrophoresis analysis confirmed the conversion of the rfaE mutant LPS to a wild-type LPS phenotype. Sequence analysis of a 2.4-kb BglII fragment revealed two open reading frames. One open reading frame encodes the RfaE protein with a molecular weight of 37.6 kDa, which was confirmed by in vitro transcription and translation, and the other encodes a polypeptide highly homologous to the *Escherichia coli* **HtrB** protein. These two genes are transcribed from the same promoter region into opposite directions. Primer extension analysis of the rfaE gene revealed a single transcription start site at 37 bp

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upstream of the predicted translation start site. The upstream promoter region contained a sequence (TA AAAT) homologous to the -10 region of the bacterial sigma 70-dependent promoters at an appropriate distance (7 bp), but not sequence resembling the consensus sequence of the -35 region was found. These studies demonstrate the ability to use complementation of defined LPS defects in members of the family Enterobacteriaceae to identify LOS synthesis genes in NTHi.

L5 ANSWER 14 OF 21 BIOSIS COPYRIGHT 1999 BIOSIS  
 AN 1995:290502 BIOSIS  
 DN PREV199598304802  
 TI Isolation and mutant analysis of the *htrb*  
 homologue of the *Haemophilus influenzae* nontypable strain  
 2019.  
 AU Lee, Na-Gyong; Sunshine, Melvin G.; Engstrom, Jeffrey; Gibson,  
 Bradford W.; Apicella, Michael A.  
 CS University Iowa, Iowa City, IA USA  
 SO Abstracts of the General Meeting of the American Society for  
 Microbiology, (1995) Vol. 95, No. 0, pp. 206.  
 Meeting Info.: 95th General Meeting of the American Society for  
 Microbiology Washington, D.C., USA May 21-25, 1995  
 ISSN: 1060-2011.  
 DT Conference  
 LA English

L5 ANSWER 15 OF 21 MEDLINE DUPLICATE 13  
 AN 93172962 MEDLINE  
 DN 93172962  
 TI The essential *Escherichia coli* *msbA* gene, a multicopy  
 suppressor of null mutations in the *htrB* gene,  
 is related to the universally conserved family of ATP-dependent  
 translocators.  
 AU Karow M; Georgopoulos C  
 CS Department of Cellular, Viral and Molecular Biology, School of  
 Medicine, University of Utah, Salt Lake City 84132.  
 NC AI21029 (NIAID)  
 GM07464 (NIGMS)  
 SO MOLECULAR MICROBIOLOGY, (1993 Jan) 7 (1) 69-79.  
 Journal code: MOM. ISSN: 0950-382X.  
 CY ENGLAND: United Kingdom  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals  
 OS GENBANK-Z11796  
 EM 199305  
 AB We report the characterization of the *msbA* gene, isolated as a  
 multicopy suppressor of the *HtrB* temperature-sensitive  
 phenotype. The *msbA* gene maps to 20.5 min on the *Escherichia*  
 Searcher : Shears 308-4994



*coli* genetic map and encodes a protein with an estimated molecular mass of 64,460 Da, with the properties of an integral membrane protein. The amino acid sequence of MsbA is very similar to those of the family of ATP-dependent translocators, which includes the haemolysin B protein of *E. coli* and the mammalian multidrug resistance (MDR) proteins. **Mutational** analysis of *msbA* indicates that it may form an operon with a downstream gene, *orfe*, and that both of these genes are essential for bacterial viability under all growth conditions tested.

L5 ANSWER 16 OF 21 MEDLINE  
 AN 93054357 MEDLINE  
 DN 93054357  
 TI The lethal phenotype caused by null **mutations** in the *Escherichia coli* *htrB* gene is suppressed by **mutations** in the *accBC* operon, encoding two subunits of acetyl coenzyme A carboxylase.  
 AU Karow M; Fayet O; Georgopoulos C  
 CS Department of Cellular, Viral, and Molecular Biology, School of Medicine, University of Utah, Salt Lake City 84132.  
 NC HL34127 (NHLBI)  
 AI21029 (NIAID)  
 GM07464 (NIGMS)  
 SO JOURNAL OF BACTERIOLOGY, (1992 Nov) 174 (22) 7407-18.  
 Journal code: HH3. ISSN: 0021-9193.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals  
 EM 199302  
 AB Insertion **mutations** in the *Escherichia coli* *htrB* gene result in the unique phenotype of not affecting growth at temperatures below 32.5 degrees C but leading to a loss of viability at temperatures above this in rich media. When *htrB* bacteria growing in rich media were shifted to the nonpermissive temperature of 42 degrees C, they continued to grow at a rate similar to that at 30 degrees C but they produced phospholipids at the rate required for growth at 42 degrees C. This led to the accumulation of more than twice as much phospholipid per milligram of protein compared with that in wild-type bacteria. Consistent with *HtrB* playing a role in phospholipid biosynthesis, one complementation group of spontaneously arising **mutations** that suppressed *htrB*-induced lethality were mapped to the *accBC* operon. This operon codes for the biotin carboxyl carrier protein and biotin carboxylase subunits of the acetyl coenzyme A carboxylase enzyme complex, which catalyzes the first step in fatty acid biosynthesis. Four suppressor **mutations** mapped to this operon. Two alleles were identified as **mutations** in the *accC* gene, the third allele was  
 Searcher : Shears 308-4994

identified as a **mutation** in the **accB** gene, and the fourth allele was shown to be an insertion of an IS1 transposable element in the promoter region of the operon, resulting in reduced transcription. The suppressor **mutations** caused a decrease in the rate of phospholipid biosynthesis, restoring the balance between the biosynthesis of phospholipids and growth rate, thus enabling **htrB** bacteria to grow at high temperatures.

L5 ANSWER 17 OF 21 MEDLINE DUPLICATE 15  
 AN 92121107 MEDLINE  
 DN 92121107  
 TI Isolation and characterization of the *Escherichia coli* **msbB** gene, a multicopy suppressor of null **mutations** in the high-temperature requirement gene **htrB**.  
 AU Karow M; Georgopoulos C  
 CS Department of Cellular, Viral and Molecular Biology, School of Medicine, University of Utah, Salt Lake City 84132.  
 NC AI21029 (NIAID)  
 GM07464 (NIGMS)  
 SO JOURNAL OF BACTERIOLOGY, (1992 Feb) 174 (3) 702-10.  
 Journal code: HH3. ISSN: 0021-9193.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals  
 OS GENBANK-M77039; GENBANK-Z11767; GENBANK-Z11768; GENBANK-L01135; GENBANK-L01136; GENBANK-L01137; GENBANK-L01138; GENBANK-L01139; GENBANK-L01140; GENBANK-L01141  
 EM 199204  
 AB Previous work established that the **htrB** gene of *Escherichia coli* is required for growth in rich media at temperatures above 32.5 degrees C but not at lower temperatures. In an effort to determine the functional role of the **htrB** gene product, we have isolated a multicopy suppressor of **htrB**, called **msbB**. The **msbB** gene has been mapped to 40.5 min on the *E. coli* genetic map, in a 12- to 15-kb gap of the genomic library made by Kohara et al. (Y. Kohara, K. Akiyama, and K. Isono, Cell 50:495-508, 1987). Mapping data show that the order of genes in the region is **eda-edd-zwf-pykA-msbB**. The **msbB** gene codes for a protein of 37,410 Da whose amino acid sequence is similar to that of **HtrB** and, like **HtrB**, the protein is very basic in nature. The similarity of the **HtrB** and **MsbB** proteins could indicate that they play functionally similar roles. **Mutational** analysis of **msbB** shows that the gene is not essential for *E. coli* growth; however, the **htrB** **msbB** double mutant exhibits a unique morphological phenotype at 30 degrees C not seen with either of the single mutants. Analysis of both **msbB** and **htrB** mutants shows that these bacteria are resistant to four

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times more deoxycholate than wild-type bacteria but not to other hydrophobic substances. The addition of quaternary ammonium compounds rescues the temperature-sensitive phenotype of **htrB** bacteria, and this rescue is abolished by the simultaneous addition of  $Mg^{2+}$  or  $Ca^{2+}$ . These results suggest that **MsbB** and **HtrB** play an important role in outer membrane structure and/or function.

L5 ANSWER 18 OF 21 MEDLINE DUPLICATE 16  
 AN 92114808 MEDLINE  
 DN 92114808  
 TI Sequencing, mutational analysis, and transcriptional regulation of the *Escherichia coli* **htrB** gene.  
 AU Karow M; Georgopoulos C  
 CS Department of Cellular, Viral, and Molecular Biology, School of Medicine, University of Utah, Salt Lake City 84132..  
 NC AI21029 (NIAID)  
 GM07464 (NIGMS)  
 SO MOLECULAR MICROBIOLOGY, (1991 Sep) 5 (9) 2285-92.  
 Journal code: MOM. ISSN: 0950-382X.  
 CY ENGLAND: United Kingdom  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals  
 OS GENBANK-X61000; GENBANK-S76453; GENBANK-X61210; GENBANK-S77306; GENBANK-S77307; GENBANK-X59272; GENBANK-X52368; GENBANK-X52369; GENBANK-X52370; GENBANK-X52371; GENBANK-X52372  
 EM 199204  
 AB The *Escherichia coli* **htrB** gene was originally discovered because its insertional inactivation led to an exquisitely temperature-sensitive phenotype in rich media, i.e. the ability to form colonies at temperatures below 32 degrees C, but not above 33 degrees C. The **htrB** gene has been sequenced. It can potentially code for two proteins, with Mr values of 35,407 Da and 8669 Da, that are encoded by overlapping, divergent open reading frames. Our data are consistent with the 35,407 Da protein being **HtrB**. Northern blot analysis clearly shows that the monocistronic **htrB** message is not under heat-shock regulation. We have also sequenced the flanking DNA and have discovered a new gene, designated orf39.9, located immediately adjacent to **htrB**, but divergently transcribed.

L5 ANSWER 19 OF 21 MEDLINE DUPLICATE 17  
 AN 91100364 MEDLINE  
 DN 91100364  
 TI Isolation and characterization of the *Escherichia coli* **htrB** gene, whose product is essential for bacterial viability above 33 degrees C in rich media.  
 AU Karow M; Fayet O; Cegielska A; Ziegelhoffer T; Georgopoulos C  
 CS Department of Cellular, Viral, and Molecular Biology, University of  
 Searcher : Shears 308-4994

Utah School of Medicine, Salt Lake City 84132.

NC AI21029 (NIAID)

GM07464 (NIGMS)

SO JOURNAL OF BACTERIOLOGY, (1991 Jan) 173 (2) 741-50.

Journal code: HH3. ISSN: 0021-9193.

CY United States

DT Journal; Article; (JOURNAL ARTICLE)

LA English

FS Priority Journals

EM 199104

AB We have identified and studied the *htrB* gene of *Escherichia coli*. Insertional inactivation of the *htrB* gene leads to bacterial death at temperatures above 33 degrees C. The mutant bacterial phenotype at nonpermissive temperatures includes an arrest of cell division followed by the formation of bulges or filaments. The *htrB+* gene has been cloned by complementation and shown to reside at 23.4 min on the *E. coli* genetic map, the relative order of the neighboring loci being *mboA-htrB-pyrC*. The *htrB* gene is transcribed in a counterclockwise fashion, relative to the *E. coli* genetic map, and its product has been identified as a membrane-associated protein of 35,000 Da. Growth experiments in minimal media indicate that the *HtrB* function becomes dispensable at low growth rates.

L5 ANSWER 20 OF 21 MEDLINE

DUPLICATE 18

AN 92021792 MEDLINE

DN 92021792

TI Complex phenotypes of null mutations in the *htr* genes, whose products are essential for *Escherichia coli* growth at elevated temperatures.

AU Karow M; Raina S; Georgopoulos C; Fayet O

CS Department of Cellular, Viral and Molecular Biology, University of Utah Medical Center, Salt Lake City 84132.

NC AI21029 (NIAID)

GM07464 (NIGMS)

SO RESEARCH IN MICROBIOLOGY, (1991 Feb-Apr) 142 (2-3) 289-94.

Journal code: R6F. ISSN: 0923-2508.

CY France

DT Journal; Article; (JOURNAL ARTICLE)

LA English

FS Priority Journals

EM 199201

AB Transposon insertion, followed by screening, has allowed the identification of a set of genes, called *htr*, whose products are required for *Escherichia coli* growth at elevated temperatures. The *htrB* gene has been shown to map at 23.5 min on the *E. coli* genetic map. It codes for a very basic, hydrophobic, 35,000-Mr polypeptide, possessing a putative

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membrane-spanning domain. At the non-permissive temperature, **htrB mutant** bacteria stop dividing, followed by the formation of bulges and eventual lysis. The **htrC** gene maps at 90 min, is under sigma 32 regulation and codes for a 21, 130-Mr polypeptide. At 43 degrees C, **htrC mutant** bacteria gradually lyse, whereas at intermediate temperatures they filament extensively. Finally, the **htrM** gene maps at 81 min, is under sigma 32 regulation and codes for a 35,000-Mr polypeptide. The **HtrM** null phenotype included inability to grow above 42 degrees C, extreme mucoidness and sensitivity to bile salts, even at the permissive temperatures. The **htrM** gene is identical to the **rfaD** gene, whose product is required for the biosynthesis of the lipopolysaccharide precursor ADP-L-glycero-D-mannoheptose (Pegues et al., J. Bact., 1990, 172, 4652-4660).

L5 ANSWER 21 OF 21 LIFESCI COPYRIGHT 1999 CSA  
 AN 91:45067 LIFESCI  
 TI Complex phenotypes of null mutations in the htr genes, whose products are essential for *Escherichia coli* growth at elevated temperatures.  
 THE BACTERIAL CELL CYCLE: STRUCTURAL AND MOLECULAR ASPECTS.  
 AU Karow, M.; Raina, S.; Georgopoulos, C.; Fayet, O.; Bouche, J.-P. [editor]; D'Ari, R. [editor]; Louarn, J.-M. [editor]  
 CS Dep. Cell., Viral and Mol. Biol., Univ. Utah Med. Cent., Salt Lake City, UT 84132, USA  
 SO RES. MICROBIOL., (1991) pp. 289-294.  
 Meeting Info.: EMBO Workshop on the Bacterial Cell Cycle: Structural and Molecular Aspects. Collonges-La-Rouge (France). 1-4 Oct 1990.  
 DT Book  
 TC Conference  
 FS J; G  
 LA English  
 SL English  
 AB Transposon insertion, followed by screening, has allowed the identification of a set of genes, called **htr**, whose products are required for *Escherichia coli* growth at elevated temperatures. The **htrB** gene has been shown to map at 23.5 min on the *E. coli* genetic map. It codes for a very basic, hydrophobic, 35,000-Mr polypeptide, possessing a putative membrane-spanning domain. At the non-permissive temperature, **htrB mutant** bacteria stop dividing, followed by the formation of bulges and eventual lysis. The **htrC** gene maps at 90 min, is under sigma super(32) regulation and codes for a 21,130-Mr polypeptide. The **htrM** gene is identical to the **rfaD** gene, whose product is required for the biosynthesis of the lipopolysaccharide precursor ADP-L-glycero-D mannoheptose.

=> d his 16-; d 1-10 bib abs

09/077572

(FILE 'CAPLUS, MEDLINE, BIOSIS, EMBASE, LIFESCI, BIOTECHDS, WPIDS, CONFSCI, SCISEARCH, JICST-EPLUS, PROMT, TOXLIT, TOXLINE, DRUGU, DRUGNL, DRUGB' ENTERED AT 10:32:59 ON 01 MAR 1999)

Author(s)

L6 969 S APICELLA M?/AU  
L7 210 S SUNSHINE M?/AU  
L8 6316 S LEE N?/AU  
L9 144 S ARUMUGHAM R?/AU  
L10 2123 S GIBSON B?/AU  
L11 3 S L6 AND L7 AND L8 AND L9 AND L10  
L12 164 S L6 AND (L7 OR L8 OR L9 OR L10)  
L13 40 S L7 AND (L8 OR L9 OR L10)  
L14 17 S L8 AND (L9 OR L10)  
L15 3 S L9 AND L10  
L16 9538 S L6 OR L7 OR L8 OR L9 OR L10  
L17 45 S (L12 OR L13 OR L16) AND L1  
L18 52 S L11 OR L14 OR L15 OR L17  
L19 10 DUP REM L18 (42 DUPLICATES REMOVED)

L19 ANSWER 1 OF 10 CAPLUS COPYRIGHT 1999 ACS DUPLICATE 1  
AN 1997:496805 CAPLUS  
DN 127:107983  
TI Non-toxic mutants of pathogenic gram-negative bacteria  
IN Apicella, Michael A.; Sunshine, Melvin G.;  
Lee, Na-gyong; Arumugham, Rasappa; Gibson,  
Bradford W.  
PA University of Iowa Research Foundation, USA; The Regents of the  
University of California; American Cyanamid Company; Apicella,  
Michael A.; Sunshine, Melvin G.; Lee, Na-Gyong; Arumugham, Rasappa;  
Gibson, Bradford W.  
SO PCT Int. Appl., 78 pp.  
CODEN: PIXXD2  
DT Patent  
LA English  
FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	WO 9719688	A1	19970605	WO 96-US18984	19961127
	W: AU, CA, JP, KR, MX, NZ, US, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM				
	RW: AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE				
	CA 2238640	AA	19970605	CA 96-2238640	19961127
	AU 9711246	A1	19970619	AU 97-11246	19961127
	EP 876150	A1	19981111	EP 96-942080	19961127
	R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, FI				
PRAI	US 95-565943		19951201		
	WO 96-US18984		19961127		
AB	A method is provided for identifying, isolating, and producing				
	Searcher : Shears 308-4994				

**htrB mutants** of gram-neg. bacterial pathogens.

The method comprises **mutating** the **htrB** gene of a gram-neg. bacterial pathogen so that there is a lack of a functional **htrB** protein, resulting in a **mutant** that lacks .gtoreq.1 secondary acyl chains contained in the wild type gram-neg. bacterial pathogen, and displays substantially reduced toxicity as compared to the wild type strain. The present invention also provides methods for using a vaccine formulation contg. the **htrB mutant**, the endotoxin isolated therefrom, or the endotoxin isolated therefrom which is then conjugated to a carrier protein to immunize an individual against infections caused by gram-neg. bacterial pathogens by administering a prophylactically effective amt. of the vaccine formulation.

L19 ANSWER 2 OF 10 CAPLUS COPYRIGHT 1999 ACS DUPLICATE 2  
 AN 1997:591347 CAPLUS  
 DN 127:288753  
 TI **Mutation of the htrB gene in a virulent**  
**Salmonella typhimurium strain by intergeneric transduction: strain**  
**construction and phenotypic characterization**  
 AU **Sunshine, Melvin G.; Gibson, Bradford W.;**  
**Engstrom, Jeffrey J.; Nichols, Wade A.; Jones, Bradley D.;**  
**Apicella, Michael A.**  
 CS Department Microbiology, University Iowa, Iowa City, IA, 52242, USA  
 SO J. Bacteriol. (1997), 179(17), 5521-5533  
 CODEN: JOBAAY; ISSN: 0021-9193  
 PB American Society for Microbiology  
 DT Journal  
 LA English  
 AB The **htrB** gene product of *Haemophilus influenzae* contributes to the toxicity of the lipooligosaccharide. The **htrB** gene encodes a 2-keto-3-deoxyoctulosonic acid-dependent acyltransferase which is responsible for myristic acid substitutions at the hydroxy moiety of lipid A .beta.-hydroxymyristic acid. Mass spectroscopic anal. has demonstrated that lipid A from an *H. influenzae* **htrB mutant** is predominantly tetraacyl and similar in structure to lipid IVA, which has been shown to be nontoxic in animal models. We sought to construct a *Salmonella typhimurium* **htrB mutant** in order to investigate the contribution of **htrB** to virulence in a well-defined murine typhoid model of animal pathogenesis. To this end, an r- m+ galeE mutS recD strain of *S. typhimurium* was constructed (MGS-7) and used in inter- and intrastrain transduction expts. with both coliphage P1 and *Salmonella* phage P22. The *Escherichia coli* **htrB** gene contg. a mini-Tn10 insertion was transduced from *E. coli* MLK217 into *S. typhimurium* MGS-7 via phage P1 and subsequently via phage P22 into the virulent *Salmonella* strain SL1344. All *S. typhimurium* transductants showed phenotypes similar to those described for the *E. coli* **htrB mutant**. Mass spectrometric anal.

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of the crude lipid A fraction from the lipopolysaccharide of the *S. typhimurium* **htrB mutant** strain showed that for the dominant hexaacyl form, a lauric acid moiety was lost at one position on the lipid A and a palmitic acid moiety was added at another position; for the less abundant heptaacyl species, the lauric acid was replaced with palmitoleic acid.

L19 ANSWER 3 OF 10 CAPLUS COPYRIGHT 1999 ACS DUPLICATE 3  
 AN 1997:731002 CAPLUS  
 DN 128:20429  
 TI Study of the role of the *htrB* gene in *Salmonella typhimurium* virulence  
 AU Jones, Bradely D.; Nichols, Wade A.; Gibson, Bradford W.; Sunshine, Melvin G.; Apicella, Michael A.  
 CS Dep. Microbiology, Univ. Iowa College Medicine, Iowa City, IA, 52242-1109, USA  
 SO Infect. Immun. (1997), 65(11), 4778-4783  
 CODEN: INFIBR; ISSN: 0019-9567  
 PB American Society for Microbiology  
 DT Journal  
 LA English  
 AB We have undertaken a study to investigate the contribution of the *htrB* gene to the virulence of pathogenic *Salmonella typhimurium*. An **htrB::mini-Tn10 mutation** from *Escherichia coli* was transferred by transduction to the mouse-virulent strain *S. typhimurium* SL1344 to create an **htrB mutant**. The *S. typhimurium* **htrB mutant** was inoculated into mice and found to be severely limited in its ability to colonize organs of the lymphatic system and to cause systemic disease in mice. A variety of expts. were performed to det. the possible reasons for this loss of virulence. Serum killing assays revealed that the *S. typhimurium* **htrB mutant** was as resistant to killing by complement as the wild-type strain. However, macrophage survival assays revealed that the *S. typhimurium* **htrB mutant** was more sensitive to the intracellular environment of murine macrophages than the wild-type strain. In addn., the bioactivity of the lipopolysaccharide (LPS) of the **htrB mutant** was reduced compared to that of the LPS from the parent strain as measured by both a *Limulus* amoebocyte lysate endotoxin quantitation assay and a tumor necrosis factor alpha bioassay. These results indicated that the *htrB* gene plays a role in the virulence of *S. typhimurium*.

L19 ANSWER 4 OF 10 CAPLUS COPYRIGHT 1999 ACS DUPLICATE 4  
 AN 1997:730328 CAPLUS  
 DN 128:21296  
 TI Evaluation of the virulence of nontypeable *Hemophilus influenzae* lipooligosaccharide **htrB** and **rfaD mutants** in the chinchilla model of otitis media

Searcher : Shears 308-4994



AU DeMaria, T. F.; Apicella, M. A.; Nichols, W. A.; Leake, E.  
R.  
CS Div. of Otologic Research, College of Medicine, Ohio State Univ.,  
Columbus, OH, 43210, USA  
SO Infect. Immun. (1997), 65(11), 4431-4435  
CODEN: INFIBR; ISSN: 0019-9567  
PB American Society for Microbiology  
DT Journal  
LA English  
AB Considerable evidence has implicated nontypeable *Hemophilus*  
*influenzae* (NTHi) lipooligosaccharide (LOS) in the pathogenesis of  
otitis media (OM); however, its exact role has not been conclusively  
established. Recently, two NTHi LOS-deficient mutants have been  
created and described. Strain 2019-DK1, an *rfaD* gene mutant,  
expresses a truncated LOS consisting of only three  
deoxy-D-manno-octulosonic acid residues, a single heptose, and lipid  
A. Strain 2019-B29, an isogenic *htrB* mutant,  
possesses an altered oligosaccharide core and an altered lipid A.  
Each strain's ability to colonize the nasopharynx and to induce OM  
subsequent to transbullar inoculation was evaluated in the  
chinchilla model. Nasopharyngeal colonization data indicate that  
the parent strain and both mutants are able to colonize the  
nasopharynx and exhibit comparable clearance kinetics. Compared  
with the parent and each other, however, the mutants demonstrated  
marked differences in virulence regarding their relative abilities  
to induce OM and persist in the middle ear post-transbullar  
inoculation. Strain B29 required a 3-log-greater dose to induce OM  
than the parent strain and did not exhibit evidence of sustained  
multiplication but persisted for the same duration as the parent.  
Conversely, strain-DK1, even when inoculated at a dose 4 logs  
greater than the parent dose, was eliminated from the middle ear 72  
h after challenge. A comparison of the relative pathogenicities of  
these isolates provides the opportunity to address fundamental  
questions regarding the contribution of LOS to pathogenesis issues  
at the mol. level. Specifically, the impact of these LOS gene  
disruptions on OM pathogenesis can be defined and may thus provide  
potential new targets for future protection and intervention  
strategies.

L19 ANSWER 5 OF 10 CAPLUS COPYRIGHT 1999 ACS DUPLICATE 5  
AN 1997:229116 CAPLUS  
DN 126:302223  
TI Identification of the ADP-L-glycero-D-manno-heptose-6-epimerase  
(*rfaD*) and heptosyltransferase II (*rfaF*) biosynthesis genes from  
nontypeable *Haemophilus influenzae* 2019  
AU Nichols, Wade A.; Gibson, Bradford W.; Melaugh, William;  
Lee, Na-Gyong; Sunshine, Melvin; Apicella, Michael A.  
CS Department of Microbiology, University of Iowa College of Medicine,  
Iowa City, IA, 52242, USA

Searcher : Shears 308-4994

SO Infect. Immun. (1997), 65(4), 1377-1386  
CODEN: INFIBR; ISSN: 0019-9567

PB American Society for Microbiology

DT Journal

LA English

AB *Haemophilus influenzae* is an important human pathogen. The lipooligosaccharide (LOS) of *H. influenzae* has been implicated as a virulence determinant. To better understand the assembly of LOS in nontypeable *H. influenzae* (NtHi), the authors have cloned and characterized the *rfaD* and *rfaF* genes of NtHi 2019, which encode the ADP-L-glycero-D-manno-heptose-6-epimerase and heptosyltransferase II enzymes, resp. This cloning was accomplished by the complementation of *Salmonella typhimurium* lipopolysaccharide (LPS) biosynthesis gene mutants. These deep rough mutants are novobiocin susceptible until complemented with the appropriate gene. In this manner, the authors are able to use novobiocin resistance to select for specific NtHi LOS inner core biosynthesis genes. Such a screening system yielded a plasmid with a 4.8-kb insert. This plasmid was able to complement both *rfaD* and *rfaF* mutants of *S. typhimurium*. The LPS of these complemented strains appeared identical to the wild-type *Salmonella* LPS. The genes encoding the *rfaD* and *rfaF* genes from NtHi 2019 were sequenced and found to be similar to the analogous genes from *S. typhimurium* and *Escherichia coli*. The *rfaD* gene encodes a polypeptide of 35 kDa and the *rfaF* encodes a protein of 39 kDa, as demonstrated by in vitro transcription-translation studies. Isogenic mutants which demonstrated truncated LOS consistent with inner core biosynthesis mutants were constructed in the NtHi strain 2019. Primer extension anal. demonstrated the presence of a strong promoter upstream of *rfaD* but suggested only a very weak promoter upstream of *rfaF*. Complementation studies, however, suggest that the *rfaF* gene does have an independent promoter. Mass spectrometric anal. shows that the LOS mols. expressed by *H. influenzae* *rfaD* and *rfaF* mutant strains have identical mol. masses. Addnl. studies verified that in the *rfaD* mutant strain, D-glycero-D-manno-heptose is added to the LOS mol. in place of the usual L-glycero-D-manno-heptose. Finally, the genetic organizations of the inner core biosynthesis genes of *S. typhimurium*, *E. coli*, and several strains of *H. influenzae* were examd., and substantial differences were uncovered.

L19 ANSWER 6 OF 10 CAPLUS COPYRIGHT 1999 ACS DUPLICATE 6

AN 1997:605036 CAPLUS

DN 127:275201

TI *htrB* of *Haemophilus influenzae*: determination of biochemical activity and effects on virulence and lipooligosaccharide toxicity

AU Nichols, W. A.; Raetz, C. R. H.; Clementz, T.; Smith, A. L.; Hanson, J. A.; Ketterer, M. R.; Sunshine, M.; Apicella, M.  
A.

CS Department of Microbiology, University of Iowa College of Medicine,  
Searcher : Shears 308-4994

Iowa City, IA, 522442, USA

SO J. Endotoxin Res. (1997), 4(3), 163-172  
CODEN: JENREB; ISSN: 0968-0519

PB Churchill Livingstone

DT Journal

LA English

AB The **htrB** mutant of *Haemophilus influenzae* (strain B29) has been shown to lack secondary (nonhydroxylated) acyl groups in its lipid A. The authors have detd. through in vitro biochem. assays that the HtrB protein acts as a specific acyltransferase in the late stages of lipid A biosynthesis and that the preferred acyl group donor is myristoyl-acyl carrier protein. Under the conditions employed, the *Escherichia coli* precursor, Kdo2-lipid IVA, functions as a myristate acceptor. Introduction of the *Haemophilus htrB* gene into an *E. coli* mutant lacking **htrB** complements the biochem. and physiol. defects assocd. with the *E. coli htrB* mutation. Tumor necrosis factor .alpha. (TNF.alpha.) assays using murine and human macrophage cells indicated that nontypeable *H. influenzae* (NtHi) strain 2019 and *H. influenzae* type b strain A2 elicit levels of expression of TNF.alpha. that are 30-40 times greater than levels induced by the isogenic **htrB** mutants (B29 and A2B29). Studies using cell-free LOS indicated that the LOS from wild type strain 2019 elicits levels of TNF.alpha. expression that are 6-8-fold higher than those of B29. In situ hybridization studies of a primary human bronchial epithelial cell line demonstrated a greater increase of TNF.alpha. message produced in the presence of 2019 LOS than in the presence of B29 LOS. TNF.alpha. levels of the cell supernatant of cells stimulated with 2019 LOS were found to be 7-8-fold higher than levels in B29 stimulated supernatants. Using the *Limulus* amoebocyte lysate for assessment of endotoxic activity, we found that wild type LOS was 8-fold higher in endotoxic activity compared with the mutant LOS. In virulence assays using i.p. inoculation of infant rats, the **htrB** isogenic strain caused bacteremia at 50% the frequency of the wild type strain. In intranasal inoculation studies, the **htrB** mutant strain was unable to cause bacteremia whereas the wild type b parent produced bacteremia in 40-60% of the animals. These findings suggest that the **htrB** gene of *H. influenzae* is important for virulence and that host TNF.alpha. expression is attenuated in response to **htrB** mutant strains.

L19 ANSWER 7 OF 10 BIOSIS COPYRIGHT 1999 BIOSIS

AN 1996:259716 BIOSIS

DN PREV199698815845

TI *Haemophilus influenzae htrB* mutants induce a reduced production of tumor necrosis factor by mouse macrophage-like cells.

AU Nichols, Wade A. (1); Sunshine, Melvin G. (1); Harty, John  
Searcher : Shears 308-4994

T. (1); Smith, Arnold L.; Apicella, Michael A. (1)  
 CS (1) Univ. Iowa, Iowa City, IA USA  
 SO Abstracts of the General Meeting of the American Society for  
 Microbiology, (1996) Vol. 96, No. 0, pp. 232.  
 Meeting Info.: 96th General Meeting of the American Society for  
 Microbiology New Orleans, Louisiana, USA May 19-23, 1996  
 ISSN: 1060-2011.  
 DT Conference  
 LA English

L19 ANSWER 8 OF 10 CAPLUS COPYRIGHT 1999 ACS DUPLICATE 7  
 AN 1995:945651 CAPLUS  
 DN 124:25327  
 TI Mutation of the *htrB* locus of *Haemophilus*  
*influenzae* nontypable strain 2019 is associated with modifications  
 of lipid A and phosphorylation of the lipo-oligosaccharide  
 AU Lee, Na-Gyong; Sunshine, Melvin G.; Engstrom,  
 Jeffery J.; Gibson, Bradford W.; Apicella, Michael  
 A.  
 CS Dep. Microbiol., Univ. Iowa, Iowa City, IA, 52242, USA  
 SO J. Biol. Chem. (1995), 270(45), 27151-9  
 CODEN: JBCHA3; ISSN: 0021-9258  
 DT Journal  
 LA English  
 AB The HtrB protein was first identified in *Escherichia coli* as a  
 protein required for cell viability at high temp., but its  
 expression was not regulated by temp. An *htrB* homolog was  
 isolated from nontypable *Haemophilus influenzae* strain (NTHi) 2019,  
 which was able to functionally complement the *E. coli htrB*  
**mutation**. The promoter for the NTHi 2019 *htrB* gene overlaps  
 the promoter for the *rfaE* gene, and the 2 genes are divergently  
 transcribed. The deduced amino acid sequence of NTHi 2019 HtrB had  
 56% homol. to *E. coli* HtrB. In vitro transcription-translation  
 anal. confirmed prodn. of a protein with an apparent mol. mass of  
 32-33 kDa. Primer extension anal. revealed that *htrB* was  
 transcribed from a  $\sigma_{70}$ -dependent consensus promoter and its  
 expression was not affected by temp. The expression of *htrB*  
 and *rfaE* was 2.5-4-fold higher in the NTHi *htrB*  
**mutant** B29 than in the parental strain. In order to study  
 the function of the HtrB protein in *Haemophilus*, 2  
 isogenic *htrB* mutants were generated by shuttle  
**mutagenesis** using a mini-Tn3. The *htrB*  
**mutants** initially showed temp. sensitivity, but they lost  
 the sensitivity after a few passages at 30.degree. and were able to  
 grow at 37.degree.. They also showed hypersensitivity to  
 deoxycholate and kanamycin, which persisted on passage. SDS-PAGE  
 anal. revealed that the lipo-oligosaccharide (LOS) isolated from  
 these mutants migrated faster than the wild type LOS and  
 its color changed from black to brown as has been described for *E.*

Searcher : Shears 308-4994

*coli htrB mutants*. Immunoblotting anal. also showed that the LOS from the *htrB mutants* lost reactivity to a monoclonal antibody, 6E4, which binds to the wild type NTHi 2019 LOS. Electrospray ionization-mass spectrometry anal. of the O-deacylated LOS oligosaccharide indicated a modification of the core structure characterized in part by a net loss in phosphoethanolamine. Mass spectrometric anal. of the lipid A of the *htrB mutant* indicated a loss of one or both myristic acid substitutions. These data suggest that HtrB is a multifunctional protein and may play a controlling role in regulating cell responses to various environmental changes.

L19 ANSWER 9 OF 10 MEDLINE DUPLICATE 8  
 AN 95172727 MEDLINE  
 DN 95172727  
 TI Molecular cloning and characterization of the nontypeable *Haemophilus influenzae* 2019 *rfaE* gene required for lipopolysaccharide biosynthesis.  
 AU Lee N G; Sunshine M G; Apicella M A  
 CS Department of Microbiology, University of Iowa, Iowa City 52242..  
 NC AI 24616 (NIAID)  
 SO INFECTION AND IMMUNITY, (1995 Mar) 63 (3) 818-24.  
 Journal code: GO7. ISSN: 0019-9567.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
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 AB The lipooligosaccharide (LOS) of nontypeable *Haemophilus influenzae* (NTHi) is an important factor in pathogenesis and virulence. In an attempt to elucidate the genes involved in LOS biosynthesis, we have cloned the *rfaE* gene from NTHi 2019 by complementing a *Salmonella typhimurium rfaE mutant* strain with an NTHi 2019 plasmid library. The *rfaE mutant* synthesizes lipopolysaccharide (LPS) lacking heptose, and the *rfaE* gene is postulated to be involved in ADP-heptose synthesis. Retransformation with the plasmid containing 4 kb of NTHi DNA isolated from a reconstituted *mutant* into *rfaE mutants* gave wild-type LPS phenotypes. Sodium dodecyl sulfate-polyacrylamide gel electrophoresis analysis confirmed the conversion of the *rfaE mutant* LPS to a wild-type LPS phenotype. Sequence analysis of a 2.4-kb BglII fragment revealed two open reading frames. One open reading frame encodes the RfaE protein with a molecular weight of 37.6 kDa, which was confirmed by in vitro transcription and translation, and the other encodes a polypeptide highly homologous to the *Escherichia coli HtrB* protein. These two genes are transcribed from the same promoter region into opposite directions. Primer extension analysis of the *rfaE* gene revealed a single

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transcription start site at 37 bp upstream of the predicted translation start site. The upstream promoter region contained a sequence (TA AAAT) homologous to the -10 region of the bacterial sigma 70-dependent promoters at an appropriate distance (7 bp), but not sequence resembling the consensus sequence of the -35 region was found. These studies demonstrate the ability to use complementation of defined LPS defects in members of the family Enterobacteriaceae to identify LOS synthesis genes in NTHi.

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TI Isolation and mutant analysis of the htrb  
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Jeffrey; Gibson, Bradford W.; Apicella, Michael A.  
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